

# Efficacy of closantel against benzimidazole resistant Haemonchus contortus infection in sheep

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#### ABSTRACT

Benzimidazoles are widely used and readily available ovine anthelminthics across the country. However, widespread resistance to this drug class has been documented, primarily in Haemonchus spp. The present study was conducted to determine the efficacy of closantel against benzimidazole resistant Haemonchus contortus infection in sheep. Naturally infected sheep (n=34) were divided into four groups on the basis of fecal egg counts (FEC) using modified McMaster technique with a lower detection limit of 8.3 eggs per gram of faeces (EPG) viz. Group-A (500-10000), Group- B (>10,000-20,000), Group-C (>20,000-30,000) and Group-D (>30,000). Generic differentiation of larvae was carried out by coproculture performed on pooled faeces which showed the presence of Haemonchus, Trichostrongylus, Oesophagostomum and Strongyloides larvae. To ascertain the evidence of benzimidazole resistance, Egg Hatch Assay (EHA) was performed. All animals were treated with Closantel @10mg\kg body weight and EPG was determined on 7th and 11th day post treatment. Animals of different groups had mean EPG range of 400-760 at 11th day of treatment. In group A, B, C and D mean per cent efficacy of closantel was 91.24±3.49, 95.15±1.72, 97.73±0.72, 98.23±0.86 respectively. Efficacy of closantel against Haemonchus and other gastrointestinal nematodes was further confirmed by performing coproculture 11th day post treatment. Post treatment coproculture revealed presence of Trichostrongylus, Oesophagostomum and Strongyloides larvae and were devoid of Haemonchus larvae. To clear the left out infection of Trichostrongylus, Oesophagostomum and Strongyloides animals were further treated with Fenbendazole @5mg/kg body weight and EPG 14 days post treatment became zero. The results of the study suggested that closantel can be used for Targeted Selective Treatment (TST) in sheep primarily infected with Haemonchus. Since closantel is highly efficacious against Haemonchus its use as an alternative to benzimidazoles group may be helpful to decrease pasture contamination. Overall control of gastrointestinal nematodiasis may therefore be possible by use of closantel along with benzimidazoles.

Key words: Benzimidazole resistance, Closantel, Efficacy, Fenbendazole, Gastro-intestinal nematodes, Sheep.

## INTRODUCTION

In India, sheep and goat farming is the main source of income to small and marginal farmers. Of about 37% of the total world's sheep population and 56% of the goat population are bred and reared in Asian countries (FAO, 2015). Parasitic gastroenteritis accounts for heavy production loss in the small ruminant industry. Infestation with these endoparasites is a severe threat to veterinary health with helminthiasis at its top (Hotez et al., 2016). Reduction in growth rates of up to 1/3<sup>rd</sup> is reported due to infestation with helminth parasites (Faizal and Rajapakse, 2001). Among all gastrointestinal nematodes that infect sheep, Family- Trichostrongylidea is predominant with Haemonchus and Trichostrongylus as major contributors to infection in animals (Rey, 1991). Infection with Haemonchus contortus is very severe (Santos et al., 2012) throughout the year with high increase in faecal egg count (FEC) in summer season (Leathwick and Besier, 2014). As a result, control of *H. contortus* is essential for ensuring both animal welfare and reducing economic losses.

Various control measures have been employed against these GI parasites which include grazing/pasture management, chemotherapy, immunoprophylaxis etc. Among these the most common and widely employed

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method under field conditions is chemotherapy. For decades, the application of broad spectrum anthelmintic has remained as primary strategy for their control. However, resistance to these anthelmintics continues to be documented in nematode populations worldwide, including India (Singh et al., 2002). Widespread resistance is present against ivermectin and fenbendazole, the most commonly available drugs against gastrointestinal nematodes (Falzon et al., 2013). Furthermore among this, most of the resistance was found in Haemonchus contortus. Development of resistant strains of Haemonchus contortus has been reported in Netherlands (Van den Brom *et al.*, 2015) where the most recently introduced amino-acetonitrile derivative (monepantel) had been used. Furthermore, reduced efficacy of the spiroindole/ML combination (derquantel/ abamectin) has been reported in Australia recently by Sales and Love (2016); (Mahieu *et al.*, 2014).

Closantel, a salicylanilide drug specifically targets haematophagus parasite such as *Haemonchus sp.* It acts by decreasing the energy level of the parasite by uncoupling oxidative phosphorylation hence deprives availability of ATP and nicotinamide adenine dinucleotide in the mitochondria (Lanusse *et al.*, 2009).

Hence, following study was carried out with the objective of determining the efficacy of closantel against GI parasites, especially *Haemonchus sp.* Moreover it was also intended to determine whether resistance was prevalent in *Haemonchus* population only or other gastrointestinal parasites too.

#### **MATERIALS AND METHODS**

#### Location of animals

Naturally infected sheep (n=34), with average age of 5-7 months were used. Animals were raised on seasonal green pastures with ad-libitum drinking water. After procurement animals were kept in semi intensive loose housing system in Division of Parasitology, Indian Veterinary Research Institute, Izatnagar, Uttar Pradesh, India (28°23'34.8°N 79°25'59.9°E). Flock was not treated with any anthelmintic expect for benzimidazoles, earlier in their life.

#### Collection of faecal samples and concentration of eggs

Faecal samples were collected from each sheep on day zero (pretreatment) and again on days 7 and 11 post closantel treatment and day 14 for fenbendazole. Samples were collected per rectally and were stored in air tight sealable plastic bags. Samples were kept in ice packs and were shifted to 4°C refrigeration in Lab until further analysis. Qualitative assessment for the presence of parasitic eggs/ ova was done by flotation method using saturated salt solution, within 48 hours.

#### Eggs per gram (EPG) of faeces by McMaster method

FEC is efficient and a cost effective method to determine parasitic burdens. Faeces were subjected to modified McMaster technique (Zajac and Conboy, 2012) and the FEC value is presented as eggs per gram (EPG). Based on the pre-treatment EPG, animals were divided into four groups: A, B, C and D.

#### Coproculture of faeces

Generic differentiation of larval types was done using Coproculture. Fresh faeces from the entire sheep flock were collected and pooled. Inverted petridish method was used along with humid conditions at 27°C for 7 days. Harvesting of L<sub>3</sub> larvae was done 7 days post incubation. Generic compositions of harvested larvae were determined as per standard keys (VanWyk and Mayhew, 2013).

## Determination of benzimidazole resistance by Egg hatch assay (EHA)

EHA was performed as per standard protocol described by Zhu et al. (2013). Eggs were exposed to different concentrations of albendazole 0.1 µg/ml, 0.3 µg/ml, 0.5 µg/ ml, 0.7 µg/ml, 1.0 µg/ml) and inhibition of hatching of larvae was determined. 0.1 µg/ml concentration of albendazole was used as discriminating dose (Coles et al., 1992). In the assay, 100 µl of egg suspension containing about 100-150 fresh eggs were dispersed into individual wells of flat bottom cell culture plates. Entire volume was made up to 500 µl with distilled water. The egg suspension in each treated well was mixed with stock albendazole to obtain the final concentration of 0.1 µg/ml, 0.3 µg/ml, 0.5 µg/ml, 0.7 µg/ml, 1.0 µg/ml. Untreated eggs in distilled water served as negative control and 1% DMSO used as emulsifier as solvent control. Plates were incubated at 27°C for 48 hours under humidified conditions. A total of hundred eggs and hatched larvae were counted under 10X of microscope. The efficacy was determined by the following equation:

Inhibition % =

Number of unhatched eggs

Number of hatched larvae + Number of unhatched eggs × 100

#### Anthelmintic treatment

Treatment-1- Closantel (ZenVet<sup>™</sup>) was given orally @10mg/ kg body to sheep having mixed infection with *Haemonchus sp*, *Trichostrongylus sp* and *Oesophagostomum sp*. as made evident by results of coproculture.

Treatment-2- Fenbendazole was given orally @5mg/kg body weight on 25<sup>th</sup> day of experiment.

Faecal samples were collected per rectally on  $7^{th}$  and  $11^{th}$  day post treatment -1 (T1) and  $14^{th}$  days post treatment-2 (T2) for FEC estimation. Efficacy of the drug was determined by using formula:

FECRT% =

(Arithmetic mean of pre-treatment EPG – Arithmetic mean of post treatment EPG)

× 100

Arithmetic mean of pre-treatment EPG

(Dash et al., 1988).

## **RESULTS AND DISCUSSION**

*Haemonchus contrortus*, a blood feeding ovine nematode is responsible for hyper acute outbreaks with FEC ranging up to >30,000 (Selemon, 2018). In the present study, based up on FEC severity, grouping of animals was done; *viz* Group-A (500-10000), Group- B (>10,000-20,000), Group-C (>20,000-30,000) and Group-D (>30,000) (Table 1). A wide variation in the range from 500-30,000 was observed. Such a high FEC ≥30,000 has been earlier reported in infection with *H. contortus* (Besier *et al.*, 2016).

Morphometric identification of the larvae through coproculture to ascertain the larval types present before

Table 1: Mean FEC with FECR of variou	is aroups.
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Parameters	Group A	Group B	Group C	Group D
Number of sheep	8	12	9	5
Mean FEC on Day 0	500-10,000	>10,000-20,000	>20,000-30,000	>30,000
Mean FEC 7 days post closantel treatment	0-1900	100-1700	0-2100	1000-2300
Mean FEC 11 days post closantel treatment	0-1100	0-3400	100-1800	100-2000
FECR, 7 days post closantel treatment	77.87 ± 10.43	94.37 ± 0.88	96.45 ± 1.11	97.68 ± 0.87
FECR, 11 days post closantel treatment	91.24 ± 3.49	95.15 ± 1.72	97.73 ± 0.72	98.23 ± 0.86
Resistance status of H. contortous	Susceptible	Susceptible	Susceptible	Susceptible
Mean FEC 14 days post fenbendazole treatment	0	0	0	0
FECR, 14 days post fenbendazole treatment	100	100	100	100

 Table 2: Generic composition of different larvae as determined by coproculture.

Anthelmintic	Gene	Generic composition of larvae			
treatment	Haemon -chus	Oesopha -gostomum	Trichost rongylus		
Pre treatment	+++	+	++		
Closantel @	Nil	+	++		
10mg/kg B.W					
Fenbendazole	Nil	Nil	Nil		
@5mg/kg B.W					

(+++ = >90%; ++ = 5-7%; + = 1-3%)

 Table 3: Egg hatch assay (EHA) for determination of benzimidazole resistance.

Albendazole concen	Inhibition of L <sub>1</sub>	Hatching of		
-tration (µg/ml)	hatching (%)	L <sub>1</sub> (%)		
0.1	36	64		
0.3	52.4	47.6		
0.5	59.4	40.6		
0.7	64.9	35.1		
1.0	70.1	29.1		

undergoing treatment showed initial mixed infection with *Haemonchus spp., Oesophagostomum spp., Trichostrongylus spp.* and *Strongyliodes spp.* with the predominance of *H. contortus.* Since majority of  $L_3$  were of *H. contortus,* this led to the formulation of hypothesis for selection of an effective drug against the blood feeding *H. contortus.* 

Simultaneously resistance to benzimidazole, a broader, widely available and most commonly used anthelmintic against gastrointestinal parasites, was checked by performing Egg Hatch Assay (EHA). The results of EHA are presented in (Table 3), which showed presence of resistance at different concentrations of albendazole. In the recent times, parasite resistant strains to benzimidazoles, macrolide lactone, monepantel, derguantel have emerged making the problem of resistance a global issue (Keane *et al.*, 2014; Cazajous *et al.*, 2018; Sales and Love (2016). Experimental design along with treatment regimen followed is presented in Fig 1 (Table 4).

Following the WAAVP guidelines which are based on FECRT, GIN infection was found susceptible to the combined treatment with closantel and albendazole (Alcalá Canto et al., 2017). Therefore, treatment with a narrow spectrum drug like closantel (ZenVet™ Oral suspension, Intas Pharmaceutical Ltd. Ahmedabad, India) @10mg/kg. particularly aiming H. contortus was undertaken. FEC post 7 and 11 days of treatment were compared which lowered significantly with a mean value of 1100, 3400,1800, 2000 in Group A,B,C,D (Table 1). A slightly higher range of FEC was noticed in Group B when compared at Day 7 and Day 11 which may be due to some concurrent infection. Similar findings have been reported in the studies of certain workers (Westers et al., 2016). Overall mean efficacy varied from 91.24% to 98.23% among different groups, after treatment. These results can be well correlated to similar observation made by previous workers (Sivajothi and Reddy, 2017), citing a 90.6% reduction in FECRT following closantel treatment in sheep.

Coproculture post closantel treatment to check reduction in infection due to *H. contortus* showed absence of *H. contortus* larvae with presence of *Oesophagostomum spp* and *Trichostrongylus spp*. Larvae were differentiated

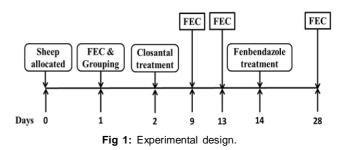
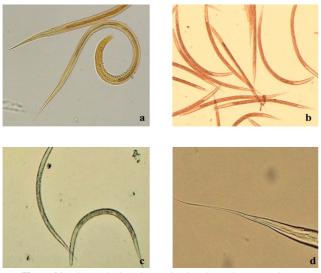


Table 4: Treatment regimen followed.

Days	Treatment	Dose rate	Average body weight	Dose volume	Route
Day 0	-	-	18.7 kg	-	-
Day 1 post 1 <sup>st</sup> FEC	Closantel (ZenVet™)	10 mg/kg Body weight	18.7 kg	5.5 ml	Oral
Day 12 post 1 <sup>st</sup> FEC	Fenbendazole (Panacur®)	5 mg/ kg Body weight	20.2 kg	4 ml	Oral



**Fig 2:** Morphometric larval examination through coproculture. (a) *Haemonchus* larvae in the coproculture (40X), (b) Predominance of *Haemonchus* in coproculture (40X), (c) Larvae of *Trichostrongylus spp.* showing pencil tail, (d) - Larvae of *Oesophagostomum spp.* with

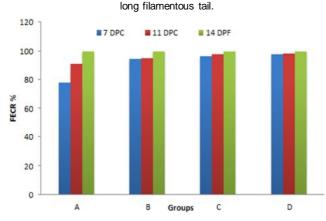


Fig 3: Percentage reductions in FEC with days.DPC: Number of days post closantel treatment; DPF: Number of days post fenbendazole treatment.

up to generic level based on morphological characteristics (Fig 2). Larval pattern during the entire study period is presented in (Table 2). To clear up the residual infection, treatment with Fenbendazole (Panacur® Vet Suspension, MSD Animal Health, Mumbai, India) @ 5mg/kg, orally was done and FEC after 14 days was determined. Post fenbendazole treatment, no parasitic eggs could be seen with 100% reduction in FEC. The combined overall efficacy of both the drugs was 100% (Fig 3). Combination of different anthelmintic classes continues to be the best approach for the efficient control of GIN as the survival of resistant genotypes is minimized (Leathwick *et al.*, 2015).

Evidently, the strain of *H. contortus* present in Indian sheep from Northern region was found susceptible to closantel with resistance to Benzimidazole group. Considering above fact, it is observed that closantel may be used to clear up very heavy infection due to *Haemonchus* in case of reported benzimidazole resistance. It is further suggested that we can use closantel in the areas where *Haemonchus* is endemic in the view of its residual effect (Anonymous, 1990). Moreover, prolonged activity of this salicylanilide has an added advantage in lowering pasture contamination, hence achieving better control (Dash, 1986). Closantel has efficacy against nematodes, trematodes and arthropods adding to its advantage for use in sheep (Maes *et al.,* 1988).

It should be kept in mind that frequent use of closantel may lead to its resistance and such an efficacious anthelmintic against *Haemonchus* may be lost in future due to indiscriminate use. Thus, to decrease the selection pressure that leads to the development of anthelmintic resistance in the animal, approach to control of nematode in the flocks must be based on diagnosis through fecal examinations, epidemiological studies and prophylaxis via nutritional strategies and targeted selective treatments (Torres Acosta *et al.*, 2012).

#### CONCLUSION

Results from the present study showed that closantel was highly effective in eliminating infection with *Haemonchus contrortus*. Therefore, in animals with mixed gastrointestinal nematodes infection, not showing response to benzimidazole treatment, infection with *Haemonchus spp*. should be confirmed and further treatment with closantel should be taken up. However, to slow down the development of anthelmintic resistance among animals, closantel should only be considered in the farms where heavy *Haemonchus* burdens are reported with documented resistance to benzimidazoles. The drug can be incorporated as a part of target strategic treatment for improved results.

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#### REFERENCES

- Alcalá Canto Y., Sumano López HS., Ocampo Camberos L and Gutiérrez L. (2017). Anthelmintic resistance status of gastrointestinal nematodes of sheep to the single or combined administration of benzimidazoles and closantel in three localities in Mexico. Veterinaria México. 3: 1-11.
- Anonymous (1990). Evaluation of certain veterinary drug residues in food. Thirty-sixth report of the joint FAO/WHO Expert Committee on Food Additives, WHO Technical Report Series, No. 799.
- Besier RB., Kahn LP., Sargison ND and Van Wyk JA. (2016). The pathophysiology, ecology and epidemiology of *Haemonchus contrortus* infection in small ruminants. Adv parasitol. 93: 95-143.

- Cazajous T., Prevot F., Kerbiriou A., Milhes M., Grisez C., Tropee A., Godart C., Aragon A and Jacquiet P. (2018). Multipleresistance to ivermectin and benzimidazole of a *Haemonchus contrortus* population in a sheep flock from mainland France, first report. Veterinary Parasitology: Regional Studies and Reports. 14:103-105.
- Coles GC., Bauer C., Borgsteede FHM., Geerts S., Klei TR., Taylor MA and Waller PJ. (1992). World Association for the Advancement of Veterinary Parasitology (WAAVP) methods for the detection of anthelmintic resistance in nematodes of veterinary importance. Vet parasitol. 44: 35-44.
- Dash KM. (1986). Control of helminthosis in lambs by strategic treatment with closantel and broadspectrum anthelmintics. Aust Vet J. 63: 4-6.
- Faizal ACM and Rajapakse RPVJ. (2001). Prevalence of coccidia and gastrointestinal nematode infections in cross bred goats in the dry areas of Sri Lanka. Small Rumin Res, 40: 233-238.
- Falzon LC., Menzies PI., Shakya KP., Jones-Bitton A., Vanleeuwen J., Avula J., Stewart H., Jansen JT., Taylor MA., Learmount J and Peregrine AS. (2013). Anthelmintic resistance in sheep flocks in Ontario, Canada. Vet parasito. 193: 150-162.
- FAO (2015). The Second Report on the State of the World's Animal Genetic Resources for Food and Agriculture, Scherf BD and Pilling D (eds). FAO Commission on Genetic Resources for Food and Agriculture Assessments, Rome (available at http://www.fao.org/3/a-i4787e/index.html).
- Hotez PJ., Pecoul B., Rijal S., Boehme C., Aksoy S., Malecela M., Tapia-Conyer R and Reeder JC. (2016). Eliminating the neglected tropical diseases: translational science and new technologies. PLoS negl tropl dis. 10: 0003895.
- Keane OM., Keegan JD., Good B., de Waal T., Fanning J., Gottstein M., Casey M., Hurley C and Sheehan M. (2014). High level of treatment failure with commonly used anthelminitics on Irish sheep farms. Irish Vet Journal. 67: 16.
- Lanusse CE, Guillermo LV and Alvarez LI (2009) Anticestodal and antitrematodal drugs. In: Veterinary Pharmacology & Therapeutics, 9<sup>th</sup> edn. [Riviere JE and Papich MG (eds)], Ames, IA: Wiley-Blackwell, 1104-1106.
- Leathwick DM and Besier RB. (2014). The management of anthelmintic resistance in grazing ruminants in Australasia-strategies and experiences. Vet Parasitol. 204: 44-54.
- Leathwick DM, Ganesh S, Waghorn TS. (2015). Evidence for reversion towards anthelmintic susceptibility in *Teladorsagia circumcincta* in response to resistance management programmes. Int J Parasitol Drugs Drug Resist. 5: 9-15.

doi: 10.1016/j.ijpddr.2015.01.001.

- Maes L., Vanparijs O and Marboom R. (1988). Abstract of the 4<sup>th</sup> Congress of the European Associationof Veterinary Pharmacology and Toxicology (Budapest), 2: 330.
- Mahieu M., Ferré B., Madassamy M and Mandonnet N. (2014). Fifteen years later, anthelmintic resistances have dramatically spread over goat farms in Guadeloupe. Vet Parasitol. 205: 379-384.
- Rey B. (1991). Small ruminant genetic resources and parasite challenge in sub-Saharan Africa.
- Sales N and Love S. (2016). Resistance of *Haemonchus* sp. to monepantel and reduced efficacy of a derquantel/abamectin combination confirmed in sheep in NSW, Australia. Vet Parasito. 228:193-196.
- Santos MC, Silva BF and Amarante AFT (2012) Environmental factors influencing the transmission of *Haemonchus contrortus*. Vet Parasito, 188: 277–284.
- Selemon M. (2018) Review on control of *Haemonchus contrortus* in sheep and goat. J Vet Med Res. 5: 1139.
- Singh D, Swarnkar CP and Khan FA. (2002) Anthelmintic resistance in gastrointestinal nematodes of livestock in India. J of Veterinary Parasito. 16: 115–130.
- Sivajothi S and Reddy BS. (2017). Therapeutic efficacy of closantel against different gastrointestinal parasites in sheep. Arch Parasitol. 1: 111.
- Torres-Acosta JF, Molento M, Fau-Mendoza de Gives P, Mendoza de Gives P. (2012) Research and implementation of novel approaches for the control of nematode parasites in Latin America and the Caribbean: is there sufficient incentive for a greater extension effort? Vet Parasitol. 186: 132-142.
- Van den Brom R., Moll L., Kappert C and Vellema P. (2015). Haemonchus contrortus resistance to monepantel in sheep. Vet Parasitol. 209: 278-280.
- VanWyk JA., Mayhew E. (2013).Morphological identification of parasitic nematode infective larvae of small ruminants and cattle: a practical lab guide. Onderstepoort. J Vet Res. 80: 1. https://doi.org/10.4102/ojvr.v80i1.539.
- Westers T., Jones-Bitton A., Menzies P., Van Leeuwen J., Poljak Z and Peregrine AS. (2016). Efficacy of closantel against ivermectin-and fenbendazole-resistant Haemonchus sp. in sheep in Ontario, Canada. Vet Parasitol. 228: 30-41.
- Zajac AM and Conboy GA (2012). Veterinary Clinical Parasitology, 8<sup>th</sup> edn. Chichester: UK, pp. 3–170.
- Zhu L., Dai JL., Yang L and Qiu J. (2013). In vitro ovicidal and larvicidal activity of the essential oil of Artemisia lancea against Haemonchus contrortus (Strongylida). Vet. Parasitol. 195: 112-117.