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# SALINITY STRESS AND SUSTAINABLE AGRICULTURE- A REVIEW

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## ABSTRACT

Soil Salinity is being recognized as a major threat to the agriculture and food production in the arid and semi-arid soils. Accumulation of sodium, magnesium and calcium salts enhances the alkalinity of the soils making the crop production difficult. Plant system has developed certain adaptation strategies for tolerating the toxicity of the salts in the soil. The present review focuses on the probable sources of soil salinity, induction of toxicity mechanisms of salinity stress tolerance in crop plants along with the strategies needed to be adapted to make agriculture a sustainable one.

Key words: Crop plants, Osmolytes, Salinity, Salinization, Sustainability, Tolerance mechanisms.

The growth and productivity of the crop plants depend on a wide variety of environmental factors such as light, water, temperature, nutrients in the soil etc. Whenever, the survival of plants is limited by these factors, than they are considered to be under stress. The most common abiotic stress factors influencing the plant productivity are salinity, drought, alkalinity, UV radiation, low and high temperatures, dust, nutrients, flood etc. Among which, salinity of the soil is a major stress factor which affects the plant growth and productivity in the semi-arid regions (Win, 2011).

Soil salinity became an aspect of serious concern to global agriculture throughout the human history (Lobell et al., 2007). Due to the extensive utilization of land, the problem became more prevalent all over the world (Egamberdieva et al., 2008; Meloni et al., 2008). Every soil possesses certain water soluble salts, which will be absorbed by the plants as plant nutrients. However, over accumulation of these salts will make the soil saline, and will suppress the plant growth. Salinization of soils is more prevalent in arid and semi arid regions. Salts are personified in the soil as ions, which will migrate upward in the soil depending on the external climatic conditions (Zhang et al., 2012). When there is dearth in the precipitation, leaching of ions from the soil profile will not takes place leading to

accumulation of salts resulting in soil salinization. This will not only effects plant growth and productivity, but also affects economic welfare, environmental health and agricultural production (Rengasamy, s2006). According to FAO Land and Nutrition Management Service (2008), over 6% of the world's land is affected by the salinity, which accounts for more than 800 million hectares of land in 100 countries with the general perception focused on arid and semi-arid regions.

Salinization is the result of two processes; 1. Primary salinization and 2. Secondary salinization. Primary salinization is the development of salinity due to natural processes such as the deposition of sand stones, alluviam in the arid and semi-arid lands. intrusion of oceans into the coastal areas followed by evapo-transpiration and high tidal intrusion of sea waterinto rivers. Certain ocean salts like Sodium chloride (NaCl) will be carried by the wind during cyclones and deposited by rainfall. Secondary process is the resultant of soil salinization by human beings as a consequence of inigation using improper methods such as poor quality of water usage, deforestration, contamination of river waters with industrial chemicals and overgrazing by domestic animals (Omami et al., 2006). Deforestration and land clearing are the major reasons for the salinization and alkalization of the soils as an

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outcome of soil migration in both the upper and lower layers (Said-Al and Omer; 2011). Anthropogenic salinization occurs in arid and semi-arid areas mainly due to water logging brought about by improper inigration (Ponnamperuma, 1984). The severity of secondary salinization arises due to poor inigration practices such as the type of system and timing and the amount of inigration water like the usage of inadequate quantities of inigration water to leach salts that accumulate in the root zone due to evaporation (Homaee *et al.*, 2002). The performance of crop plants in salinized soils is not only affected by the salinity levels but also by the soil characteristics (Burger and Èelková 2003).

Types of salts active in soil salinization: Salts of several cations such as sodium (Na<sup>+</sup>), magnesium  $(Mg^{2+})$ , calcium  $(Ca^{2+})$  are found to induce salinity to soil, however, sodium is the most dominant salt causing detenioration of the physical structure of the soils. Sodicity is the result of secondary salinization in day soils, where leaching occurs through human induced processes which washes soluble salts into the subsoil and leaves sodium bound to the negative charges of the clay due to an increase in its concentration (Sangeeta et al., 2011). The common anions are chloride (Cl), sulphate (SO<sup>2</sup>) and carbonate (HCO, ). Both Nat and Cl are toxic to plants (Hasegawa et al., 2000). Historically soils were classified as saline, sodic or salinesodic based on the total concentration of salt and the ratio of Na<sup>+</sup> to Ca<sup>2+</sup> and Mg<sup>2+</sup> in the saturated extract of the soil (Dubey, 1997).

Nitrogen fertilizers and nitrate leaching: Nitrogen fertilizers such as usea and ammonium nitrate have a high leaching potential and mobility. During the past few years excessive use of the ammonia and usea fertilizers resulted in leaching of soils below the root zone, contaminating groundwater. Fertilizers misused for higher yield is an important cause of nitrate contamination. Other agents such as animal manue, domestic waste water and *in situ* organic matter deposition in soils are exacerbating the risk. Crop management and nitrogen fertilization techniques are important in reduction of nitrogen losses since annual crop's nitrogen uptake efficiency is < 50% and about 30–50% of applied nitrogen fertilizer is leached into groundwater (Ventura *et al.*, 2008). Interactive effect of nitrogen fertilizers and salinization: Nutrient deficiency limits plant growth under low salinity, which results in increased salt tolerance. In moderate or high saline conditions, increased concentration of salts hinders the growth. In most of the saline soils, application of fertilizers enhanced the soil salinity, which intum increased the yield and productivity of cotton, millet, rice, com and wheat. Plants will reduce their tolerance to salt when nitrogen is applied to extreme saline soils. This might be the reason for similitude behavior of both halophytes and glycophytes in high and mild saline soil conditions coupled with deficiency of nitrogen. However, biomass of some plants like cotton and com were found to be increased when nitrogen is applied in less saline conditions. High concentrations of salt ions, usually Na and Cl affect the activity of nutrients in the soil, resulting in a nutritional disorder in plants. Nutrient status in plant tissues may be associated with the interactive effect of salinity and nutrients on the plant growth. Nutrient uptake and accumulation by plants was affected under saline conditions due to the competition among various salt species which also further affected by composition of soil (Khorsandi and Anagholi, 2009). Nat induces deficiency of Kt during salinity stress although plants selectively absorb potassium over sodium.

Impact of soil salinization in India: In India increasing soil salinity and its damaging effects on agricultural soils severely affecting the agriculture yield, which has become not only an environmental concern but also an economical issue. Year after year increased salinization of agricultural land is reported in the sub-continent due to accumulation of salts in the soil. Among the 9.38 million hectares of salinized soils in India, 3.88 million hectares are alkali soils and 5.5 million hectares are salinized due to the climatic and environmental factors (Jaleel et al., 2008). Soil salinization increases in proportion due to lack of rainfall and inegular farming practices. In Indian sub-continent, average rainfall is approximately 1100mm, out of which 46% of rain water merges into various water bodies. 32 % water gets evaporated and only 22% waterfrom the rainfall recharges the ground aquifers and the soil. Even though this water resource is huge in nature it distributes across India seasonally and spatially due

to vagaries of monsoon, resulting in floods or drought. Floods result in the runoff of the water excessively, through rivers into Bay of Bengal or Arabian Sea, leading to a process of natural salinization due to the wash of soil top layers and accumulation of salts mostly in the coastal areas of the sub-continent (Hemapriya *et al*, 2010).

Arid and semi-arid regions of India and the remaining world are more vulnerable to salinity risk, which is found to be a serious threat to the agriculture effecting both growth as well as the yield. Legume grains have very less defiance to the salinity stress since their levels of tolerance are less to salinity and more over their symbiotic nitrogen fixation is sensitive when higher salinity rates predominates the area (Ramana et al., 2012). Rhizobial infection and root nodule development are particularly sensitive to salinity. Even though soil salinization does not affect the rhizobial activity in colonization but will reduce the efficacy of the fully grown nodules which are developed prior to the saline conditions. Apart from these ill effects, saline conditions hamper the growth of newly initiated nodules (Rao et al., 2002). Studies on cereals indicates that salinity stress decreased relative water content, chlorophyll content, carotenoids, biomass and grain yield. (Sai Ram et al, 2002).

Agriculture in arid and semi-arid conditions also enhanced the salinity of the soils. Indian farmers are depending on the ground water for their agriculture needs resulting in the decrease of permanent water table levels leading to the accumulation of salts at the root zone. Enhanced salinity and sodicity of the soils will have adverse effect on the physical structure of the soil which intum resulting in decreased aeration and water transfer However; farmers are attempting to resolve this problem by inigating their farm lands either with a potpouni of canal water and ground water or alternate inigation (Dutta *et al.*, 2002)

## **Effect of Soil Salinization**

Agriculture: One of the main factors exacerbating the rate of salinization in most of the inigated soils is the poor drainage system and availability of ample inigation water (Town *et al.*, 2008). During the general process of water absorption by plants or due to evaporation, salts in the water will remain in the soil, contributing to the soil salinization. This process

along with the improper inigation management is increasing the risk of soil salinization day by day. One of the prominent methods to prevent this is to pass surplus amount of water through the root zone in order to leach the salts from soil (Maas et al. 1999). However, some of these salts are essential to plants as plant nutrients and some salts such as nitrates act as pollutants if leached into groundwater in excess. Osmotic effect is the first outcome of salinization of the soil requiring an additional energy input of plants to absorb water, which means that the soil need to be moist in order to supply similitude amount of water to plant. More energy is exerted by the plant due to osmotic effect caused by soil salinization whereas not essential under normal conditions (Romero et al., 2001). This energy can otherwise be used for the purpose of flowering, finiting or growth. When threshold tolerance exceeds, plant's growth is affected resulting in its death due to excess salt concentration (Sohan et al., 1999).

Yield of crop plants: Salinization severely affects the agricultural productivity of crop plants. The salinization of agricultural lands at massive scale causes severe economic loss at the global level. The annual global income losses due to salinization of agricultural land were \$11.4 billion in inigated land and \$1.2 billion in non-irrigated areas (Zhu et al., 2005). Nutritional imbalance coupled with excessive salt in the leaves will affect the photosynthetic system causing chlorosis and death of plant in saline soils (Munns and Tester, 2008). One of the limiting factors for plant survival in saline soils is leaf senescence (Parvaiz and Satyavati, 2008). In saline soils, plant density is an important factor which affects the salinity in crop plants. However, optimum plant density determination is highly difficult since levels of salinity and stress tolerance of several crop plants varies. This strategy may work in reducing the impact of salinity on plants of a field but cannot improve the yield since vegetative growth is more favored than crop yield (Ali et al., 2009). Soil Salinization decreases shoot growth, germination and seed yield with low fiber quality of cotton crop (Dong et al., 2009; Higbie et al., 2010). Rice crop in the coastal regions of Asia is frequently affected by exposure to sea water brought in by cyclones (Sultana et al., 2001). Rice (Oryza sativa) is one of the most sensitive and barley (Hordeum vulgare) is the most tolerant cereal to the soil salinity, whereas

Bread wheat (*Thiticum aestivum*) is moderately tolerant. Tall wheatgrass (*Thinopyrum ponticum*, syn. *Agropyron elongatum*) is one of the most tolerant member of salinity among the monocots, where its growth proceeds in the concentrations of salt as high as in seawater (Munns, 2005). The variation in salinity tolerance of dicotyledonous species is even greater than in monocotyledonous species. Salt treatment caused a significant decrease in relative water content (RWC) in sugarbeet varieties (Ghoulam *et al.*, 2002). Legumes are very sensitive, to salinity. Alfalfa (*Medicago sativa*) is highly tolerant along with halophytes such as saltbush (*Atriplex* spp.), which grows well at salinities greater than that of seawater (Mariam *et al.*, 2006).

Response of plants to salinity stress: Salinity is an increasing threat to many inigated, and and semiarid areas of the world, accomplished by the insufficient atmospheric precipitations to leach the salts from the root zone adversely affecting the crop productivity (Neumann, 1997). The performance of the plants in saline conditions also depends on the, environmental conditions such as humidity, temperature, light, growth stage, sensitivity of the plants etc (Munns, 2002). Effects of salinity on plants are similar to that of water deficiency, where long term exposure will have specific affects, such as reduced rate of leaf emergence and heavier impact on leaves than on roots. Extended exposure of plants to salts for months leads to death of whole plant before seed maturation (Maggio et al., 2011).

Photosynthesis and antioxidant defense system: Primary response of crop plants to the increased amounts of salts is stunted growth (Romero-Aranda et al., 2001). Accumulation of salts in the leaves causes premature aging, reduces the nutrient supply and products of photosynthesis thus impairing the growth of the entire plant. In salt sensitive genotypes, the accumulation of salts will lead to their death. Photosynthesis is extremely sensitive to salinity stress. Accumulation of salts causes an irreparable damage to the photosynthetic apparatus due to dehydration of cell membranes and closure of stomata which reduces their permeability to CO. (Piotr et al., 2009). High concentrations of salts (above 250 mM) will cause metabolic limitations of photosynthesis in leaves (Mums et al., 2006). Genes involved in the photosynthetic pathways are not much altered by the salinity stress in stress tolerant plant *Thellungiella* (Wu *et al.*, 2012), while in rice alterations in photosynthesis related genes are mostly associated with stress recovery (Zhou *et al.*, 2009). High salt concentrations in the soil will trigger many changes in the plants, leading to their recovery and enhanced tolerance, which are less understood. Facultative halophyte *Mesembryanthemum crystallimm* uses CAM pathway under high salinity concentrations instead of the usual C3 pathway (Cushman *et al.*, 2008). Another plant *A triplex lentiformis* is salt tolerant due to the property that the C4 pathway runs along instead of C3 biochemical pathway (Zhu and Meinzer, 1999).

One of the byproducts of the vital photosynthetic reactions taking place in the plants is the reactive oxygen species (ROS), whose production is enhanced under abiotic stress causing ineparable damage to the cells (Candan and Tarhan, 2003). The reduced rate of photosynthesis increases the formation of reactive oxygen species ROS (Foyer and Noctor, 2005). Basically, ROS are partially reduced forms of atmospheric oxygen. In order to produce water in these processes, four electrons are required for the reduction of oxygen. But the transference of one, two and three electrons results in the formation of ROS respectively, to 0, to form superoxide (0, ), hydrogen peroxide (H, 0,) and hydroxyl radical (HO<sup>-</sup>). These species of oxygen are highly toxic and reacts with biomolecules such as lipids, proteins, nucleic acid, etc. causing lipid peroxidation, protein denaturing and DNA mutations (Seyed et al., 2011).

The plants have developed defense mechanisms comprising of enzymatic and nonenzymatic antioxidant system (Ramana et al., 2012). The alleviation of oxidative damage and increased resistance to environmental stresses is often correlated with an efficient antioxidative system comprising enzymatic antioxidants such as superoxide dismutase, (SOD), catalase (CAT), ascorbate peroxidase (APX), glutathione reductase (GR) and monodehydroascorbate reductase (MDAR) and non-enzymatic antioxidants glutathione, ascorbate and carotenoids whose concomitant effort will reduce the toxicity of the ROS (Attia et al., 2009). Activity of Mn-SOD was enhanced under salt stress in wheat, while Cu/Zn-SOD activity remained constant (Hernandez et al.,

1999). Under elevated levels of SOD, scavenging of superoxide radical will be properly done, minimizing the damage to membranes there by leading to the increased tolerance to oxidative stress (Mider, 2002). Over expression of genes leading to increase in the activities of Mn-SOD, Fe-SOD, Cu/Zn-SOD, bacterial catalase and glutathione-S-transferase (GST)/glutathione peroxidase (GPX) can increase the performance of plants under salinity stress. In leaves of tice plant, salt stress enhanced the contents of H<sub>2</sub>O<sub>2</sub> and the activities of APX and GPX (Lee *et al.*, 2001). Tomato plants under high salinity concentrations showed higher antioxidant enzyme activities (Rodriguez-Rosales *et al.*, 1999).

Accumulation of osmolytes and proteins: Plants adapt to salinity through maintaining internal water potential below that of soil and turgor pressure for uptake of water by synthesizing and accumulating low molecular weight compatible osmolytes (Tester and Davenport, 2003). During osmotic stress, cytoplasm synthesizes osmolytes because they are inert to the biochemical reactions and they replace water. Majority of the osmolytes are organic solutes consisting of simple sugars (glucose and fructose), sugar alcohols (glycerol), complex sugars (Trehalose), quarternary amino acid derivatives (proline and glycine betaine), sulfonium compounds (cholineo-sulfate and dimethyl sulfonium propironate) with a very few exceptions like K<sup>+</sup> ion (Yokoi et al., 2002). The accumulation of carbohydrates such as glucose, fructose, sucrose, fructans has been widely reported in plants as a response to salinity stress, playing a major role in osmoprotection, osmotic adjustment, carbon storage and radical scavenging (Nasrin et al., 2010). Trehalose, a suppressor of apoptotic cell death accumulates under abiotic stresses like salinity, protects membranes and proteins in cells and reduces the aggregation of denatured proteins (Yamada et al., 2003). Proline is a very active compatible osmolyte, whose accumulation contributes to the membrane stability and mitigates the effect of NaCl on cell membrane disruption (Farouk, 2011). Transgenic tobacco plants, which induces over production of proline showed tolerance to the NaCl (Hong et al., 2000). Glycine betaine is a small organic water soluble

non-toxic metabolite that plays a crucial role in effective protection against salt stress (Chen and Murata, 2008). The major role of glycine betaine is protecting plant cells from salt stress by osmotic adjustment, protein stabilization, photosynthetic apparatus protection and reduction of oxygen radical scavengers (Suriyan and Chalempol, 2010). Glycine betaine is reported to get accumulated in response to osmotic stress in many crops, including spinach, barley, tomato, potato, rice, carrot and sorghum (Yang *et al.*, 2003).

Acclimation of plants to environmental factors like salinity will occur by internal adjustments within the tissues and cells, enabling the plant metabolism to proceed under these conditions. In contrast to adaptation that occurs in plant phylogeny, acclimation occurs during plant ontogeny, describing enhanced stress tolerance of a particular individual, which was widely reported in many plant species with increasing ability to tolerate salt stress after being exposed to a brief period of time (Djanaguiraman et al., 2006). The acclimation of plants to salinity will benefit them agronomically by improved survival rate, higher growth rate with less biomass reduction and physiologically by lowering the Na<sup>+</sup> accumulation in the shoot, facilitating better osmotic adjustment (Camila et al., 2012). However, the plant protection and acclimation mechanisms beyond this acquired resistance to salinity remain largely a matter of conjecture.

Conclusions and future perspectives: Soil salinity is causing huge amount of damage to the agriculture and crop productivity globally, conventional approaches are being followed using the available information in plant physiology, genetics and biochemical methodologies for studying plant responses to abiotic stresses have begun to bearfiuit in order to combat with the toxicity generated by the salinity in soils. Transgenic plants with enhanced tolerance to the salinity stress were generated, whose productivity in the field and price is yet to be stabilized.

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### REFERENCES

- Ali, H., Afzal, M.N. and Muhammad, D. (2009). Effect of sowing dates and plant spacing on growth and dry matter partitioning in cotton (*Gossypium hisutum* L). *Pak J. Bot.*, 41: 2145–2155.
- Attia, H., Kanay, N. and Lachaâl, M. (2009). Light interacts with salt stress in regulating superoxide dismutase gene expression in *Arabidopsis. Plant Sci.*, 177: 161-167.
- Burger E and Celková, A. (2003). Salinity and sodicity hazard in water flow processes in the soil. *Plant Soil Environ.*, 49: 314–320.
- Camilla, P., Stefano, M. and Sergey, S. (2012). Physiology of acclimation to salinity stress in pea (*Pisum sativum*). *Environ. Exp. Bot.*, 84: 44 – 51.
- Candan, N. and Tahan, L.(2003). The correlation between antioxidant enzyme activities and lipid peroxidation levels in *Mentha pulegium* organs grown in Ca<sup>2+</sup>, Mg<sup>2+</sup>, Cu<sup>2+</sup>, Zn<sup>2+</sup> and Mn<sup>2+</sup> stress conditions. *Plant Sci.*, 163: 769-779.
- Chen, T.H.H. and Murata, N. (2008). Glycinebetaine: an effective protectant against abiotic stress in plants. *Trend Plant Sci.*, 13: 499-505.
- Cushman, J.C., Agarie, S., Albion, R.L., Elliot, S.M., Taybi, T. and Borland, A.M. (2008). Isolation and characterization of mutants of common ice plant deficient in Crassulacean Acid Metabolism. *Plant Physiol.*, 147: 228–238.
- Datta, K.K., Jong, C.de., (2002) Adverse effect of water logging and soil salinity on crop and land productivity in northwest region of Haryana, India, *Agricultural water management*, 3:223–238.
- Djanaguiraman, M., Sheeba, J.A., Shanker, A.K., Devi, D.D. and Bangarusanny, U. (2006). Rice can acclimate to lethal level of salinity by pretreatment with sublethal level of salinity through osmotic adjustment. *Plant and Soil*, 284: 363–373.
- Dong H. Z., Li, W.J., Tang, W. and Zhang, D.M. (2009). Early plastic mulching increases stand establishment and lint yield of cotton in saline ûelds. *Field Crops Res.*, 111: 269–275.
- Dubey, R.S. (1997). Photosynthesis in plants under stressful conditions. In: Handbook of Photosynthesis.c Pessarakli, M. (eds) *Marcel Delder:* New York. USA. 859-975.
- Egamberdieva, G.M. E., Albacete, A Martý´nez-Andujar, C., Acosta, M., Romero-Aranda, R., Dodd, I.C., Lutts, S. and Pe¢rez-Alfocea, E (2008). Hormonal changes during salinity-induced leaf senescence in tomato (*Solarum lycopensicum* L.). *J. Exp. Bot.*, 59: 3039–3050.
- Farouk, S. (2011). Osmotic adjustment in wheat flag leaf in relation to flag leaf area and grain yield per plant. *Journal of Stress Physiology & Biochemistry*, 7: 117-138.
- Foyer, C.H. and Noctor, G. (2005). Oxidant and antioxidant signaling in plants: a re-evaluation of the concept of oxidative stress in a physiological context. *Plant Cell Environ*.28: 1056–1071.
- Ghoulam, C., Foursy, A. and Fares, K. (2002). Effects of salt stress on growth, inorganic ions and proline accumulation in relation to osmotic adjustment in five sugar beet cultivars. *Environ. Exp. Bot.*, 47: 39-50.
- Hasegawa, PM., Bresan, R.A., Zhu, J.K. and Bohnert, H.J. (2000). Plant cellular and molecular responses to high salinity. Ann Rev. Plant Physiol. Plant Mol. Biol., 51: 4632-4649.
- Hemapriya, R., Sankar K. and Imran A. Dar (2010). Geologic and geomorphologic investigation of Gadilam riverbasin (India). *Journal of Environmental Research and Development*, 4: 3.
- Hernandez, J.A., Campillo, A., Jimenez, A., Alacon, J.J. and Sevilla, E (1999). Response of antioxidant systems and leaf water relations to NaCl stress in pea plants. *New Phytol*, 141: 241–251.
- Higbie, S. M., Wang, E., Mc, D., Stewart, J., Sterling, T.M., Lindemann, W.C., Hughs, E. and Zhang, J. (2010). Physiological response to salt (NaCl) stress in selected cultivated tetraploid cottons. *Int. J. Agron.*, 1: 1–12.
- Homaee, M., Feddes, R.A. and Dirksen, C. (2002). A macroscopic water extraction model for nonuniform transient salinity and water stress. *Soil Sci. Soc. Am. J.*, 66: 1764–1772.
- Hong, Z., Lakkineni, K., Zhang, Z. and Venma, D.P.S. (2000). Removal of feedback inhibition of 1-pynoline-5-carboxylate synthetase results in increased proline accumulation and protection of plants from osmotic stress. *Plant Physiol*, 122: 1129–1136.
- Jaleel, C.A., Sankar, B., Suidharan, R., Paneerselvam, R. (2008). Soil Salinity Alters Growth, Chlorophyll Content, and Secondary Metabolite Accumulation in *Catharanthus roseus*, *Turkish Journal of Biology*, 32: 79-83.
- Khorsandi, E and Anagholi, A. (2009). Reproductive compensation of cotton after salt stress relief at different growth stages. J. Agron. Crop Sci., 195: 278–283.
- Lee, D.H., Kim, Y.S. and Lee, C.B. (2001). The inductive responses of the antioxidant enzymes by salt stress in the rice (*Oryza sativa*). *J. Plant Physiol* 158: 737–745.
- Lobel, D. B., Ortiz-Monsterio, J.I., Gunola, EC. and Valenzuuela, L. (2007). Identification of saline soils with multiyear remote sensing of crop yields. Soil Sci. Soc. Am. J., 71: 777–783.

- Maas, E.V. and Grattan, S.R. (1999). Crop yields as affected by salinity. In: Handbook of Photosynthesis. Pessaraki, M. (eds). *Marcel Delder:* New York. USA. 55–108.
- Maggio, A., De Pascale, S., Fagnano, M. and Barbieri, G. (2011). Saline agriculture in Mediterranean environments. *Italian journal of Agronomy*, 6: 36-43,
- Mariam, K., Noel, A.T. and Carmen L. (2006). Alleviation of salt stress in common bean (*Phaseolus vulgaris*) by exogenous abscisic acid Supply. *J. Plant Growth Regul*, 25: 110–119.
- Meloni, D. A., Oliva M.A., Martinez, C.A. and Cambraia, J. (2003). Photosynthesis and activity of superoxide dismutase, peroxidase and glutathione reductase in cotton under salt stress. *Environ. Exp. Bot.*, 49: 69–76.
- Mittler; R. (2002). Oxidative stuess, antioxidants and stuess tolerance, Thends in Plant Science, 7: 405-410.
- Munns, R. (2002). Comparative physiology of salt and water stress *Plant Cell Environ.*, 25: 239–250.
- Munns, R. (2005). Genes and salt tolerance: bringing them together: New Phytol, 167: 645-656.
- Munns, R., James, R.A. and La<sup>\*\*</sup>uchli, A. (2006). Approaches to increasing the salt tolerance of wheat and other cereals. *J. Exp. Bot.*, 57: 1025–1043.
- Munns, R. and Tester, M. (2008). Mechanisms of salinity tolerance. Annu. Rev. Plant Biol., 59: 651–668.
- Nasnin, A, B., Anamul, H., Megumi, W-S., Mohammad, M.I., Misugi, U., Ken, M., Yoshimasa, N. and Yoshiyuki M. (2010). Proline and glycine betaine Ameliorated NaCl Stress via Scavenging of hydrogen peroxide and methyl glyoxal but not superoxide or nitric oxide in tobacco cultured cells. *Bioscience, Biotechnology and Biochemistry*, 74: 2043-2049.
- Neumann, P (1997). Salinity resistance and plant growth revisited. Plant, Cell Environ., 20: 1193-1198.
- Omani, E.N., Hammes, P.S. and Robbertse, P.J. (2006). Differences in salinity tolerance for growth and water use efficiency in some amaranth (*Amaranthus* spp.) genotypes. *NewZeal J. Crop Hort Sci.*, 34: 11-22.
- Parvaiz, A. and Satyavati, S. (2008). Salt stress and phyto-biochemical responses of plants A review. *Plant Soil Environ.*, 54: 89-99.
- Piotr, S. and Giles N.J. (2009). Contrasting responses of photosynthesis to salt stress in the glycophyte Arabidopsis and the halophyte Thellungiella: Role of the plastid terminal oxidase as an alternative electron sink. Plant Physiol, 149: 1154-1165.
- Ponnamperuma, EN. (1984). Role of cultivar tolerance in increasing rice production in saline lands. In: Salinity tolerance in plants: strategies for crop improvement. [Staples, R.B. and G.H. Toenniessen (eds)]. John Wiley and Sons, New York 255-271.
- Ramana, G.V., Sweta Padma, P. and Chaitanya, K.V. (2012). Differential responses of four soybean (*Glycine max* L.) cultivars to salinity stress. *Legume Res.*, 35: 185 193.
- Rao, D.L.N., Giller, K.E., Yeo, A.R., Flowers, T.J. (2002). The effects of salinity and sodicity upon nodulation and mitrogen fixation in chickpea (*Cicer anietinum*). *Annals of Botany* 89:563-570
- Rengasamy, P (2006). World salinization with emphasis on Australia. J. Exp. Bot., 57: 5, 1017–1023.
- Rodriguez-Rosales, M.P., Kerkeb, L., Bueno, P and Donaire, J.P. (1999). Changes induced by NaCl in lipid content and composition, lipoxygenase, plasma membrane H<sup>+</sup> ATPase and antioxidant enzyme activities of tomato (*Lycopeusicon esculantum* Mill.) cali. *Plant Sci.*, 143: 143–150.
- Romero, A., Soria R.T. and Cuartero, S. (2001). Tomato plant-water uptake and plant-water relationships under saline growth conditions. *Plant Sci.*, 160: 265-272.
- Romero-Aranda, R., Soria, T. and Cuartero, J. (2001). Tomato plant water uptake and plant water relationships under saline growth conditions. *Plant Sci.*, 160: 265–272.
- Said-Al, A. and OmerE.A. (2011). Medicinal and aromatic plants production under salt stress. A review. *Herba Polonica*, 57: 72-87.
- Sai Ram, R., Veerabhadra Rao, K., Srivastava, G.C. (2002). Differential response of wheat genotypes to long term salinity stress in relation to oxidative stress, antioxidant activity and osmolyte concentration, *Plant science*, 5: 1037–1046.
- Sangeeta, Y., Irfan, M., Aqil, A. and Shamsul, H. (2011). Causes of salinity and plant manifestations to salt stress: A review. J. Environ. Biol., 32: 667-685.
- Seyed, M., Shahab, J., Bahram, A., Mohammad, Z., Rasool, A.Z. and Majid, K. (2011). Soil salinity alters the morphology in *Catharanthus roseus* and its effects on endogenous mineral constituents. *Middle-East Journal of Scientific Research*, 7: 07-11.
- Sohan, D., Jason, R. and Zajcek, J. (1999). Plant-water relations of NaCl and calcium-treated sunflower plants. *Env. Exp. Bot.*, 42: 105-111.

- Sultana, N., Ikeda, T. and Kashem, M.A. (2001). Effect of foliar spray of nutrient solutions on photosynthesis, dry matter accumulation and yield in seawater stressed rice. *Environ. Exp. Bot.*, 46: 129-140.
- Suriyan, C. and Chalempol, K. (2010). Effect of glycine betaine on proline, water use, and photosynthetic efficiencies, and growth of rice seedlings under salt stress. *Turk J. Agric. For.*, 34: 517-527.
- Tester; M. and Davenport R. (2003). Nat tolerance and Nat transport in higher plants. Ann. Bot., 91: 503–527.
- Town, M. H., Chandrasekhar, T., Mahamed, H., Zafar, S., Brhan, K.S. and Rama Gopal, G. (2008). Recent advances in salt stress biology – a review. *Biotechnology and Molecular Biology Review*, 3: 8-13.
- Ventura, M., Scandellari, F., Ventura, F., Guzzon, B., Rossi P.P. and Tagliavini, M. (2008). Nitrogen balance and losses through drainage waters in an agricultural watershed of the Po Valley (Italy). *Europ. J. Agronomy*, 29:108-115.
- Win, K.T. (2011). Genetic analysis of Myanmar *Vigna* species in responses to salt stress at the seedling stage. *Afr. J. Biotechnol*, 10: 1615-1624.
- Wu, H.J., Zhang Z., Wang J.Y., Oh, D.H., Dasanayake, M., Liu, B., Huang Q., Sun, H.X., Xia, R., Wu, Y., Wang Y.N., Yang Z., Liu, Y., Zhang W., Zhang H., Chu, J., Yan, C., Fang S., Zhang J., Wang Y., Zhang E, Wang G., Lee, S.Y., Cheeseman, J.M., Yang B., Li, B., Min, J., Yang L., Wang J., Chu, C., Chen, S.Y., Bohmert, H.J., Zhu, J.K., Wang X.J. and Xie, Q. (2012). Insights into salt tolerance from the genome of *Thelungiella salsuginea*. Proc. Natl Acad. Sci USA., 109: 12219-12224.
- Yamada, T., Takatsu, Y., Manabe, T., Kasumi, M. and Marubashi, W. (2003). Suppressive effect of trehalose on apoptotic cell death leading to petal senescence in ethylene-insensitive flowers of gladiolus. *Plant Sci.*, 164: 213–221.
- Yang, W.J., Rich, P.J., Axtell, J.D., Wood, K.V., Bonham, C.C., Ejeta, G., Mickel, B.M.V. and Rhodes, D. (2003). Genotypic variation for glycinebetaine in sorghum. *Crop Sci.*, 43: 162–169.
- Yokoi, S., Quintero, E.J., Cubero, B., Ruiz, M.T., Bressan, R.A., Hasegawa, P.M. and Pardo, J.M. (2002). Differential expression and function of *Arabidopsis thaliana* NHX Na<sup>+</sup>/H<sup>+</sup> antiporters in the salt stress response. *Plant J.*, 30: 529–539.
- Zhang, H.J., Dong, H.Z., Li, W.J. and Zhang, D.M. (2012). Effects of soil salinity and plant density on yield and leaf senescence of field-grown cotton. Agronomy & Crop Science, 198: 27–37.
- Zhou, Y.H., Wu, J.X., Zhu, L.J., Shi, K. and Yu, J.Q. (2009). Effects of phosphorus and chilling under low irradiance on photosynthesis and growth of tomato plants. *Biol. Plant.*, 53: 378-382.
- Zhu, J. and Meinzer, E.C. (1999). Efficiency of C4 photosynthesis in A triplex lentiformis under salinity stress. A ustral. J. Plant Physiol., 26: 79–86.
- Zhu, J.K., Bressan, R.A., Hasegawa, P.M., Pardo, J.M. and Bohmert, H.J. (2005). Salt and crops: salinity tolerance. News CASE News from the Council for Agricultural Science and Technology, 32: 13–16.