



# Ruling out Efficacy of Hormonal Protocols alone or Fortified with Mineral and Bypass Fat for Treatment of Postpartum Anoestrus in Cattle

Chandra Prakash Dixit<sup>1</sup>, D. Bhuyan<sup>1</sup>, M. Bhuyan<sup>1</sup>, K. Ahmed<sup>1</sup>,  
A. Baruah<sup>2</sup>, D. Kalita<sup>3</sup>, S. Haloi<sup>4</sup>, M. Malik<sup>5</sup>

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## ABSTRACT

**Background:** High milk production and excellent fertility is a dream of livestock owners. The major reproductive disorders which lead to decrease the productivity and reproductive ability of farm animals is postpartum anestrus. Recently, kisspeptin was reported as game changer in the animal reproduction. Kisspeptin increases the number of follicles at wave emergence, oestrus response rate and duration of oestrus. Thus, indicating its potentiality for induction of oestrus in anestrus animals.

**Methods:** The present study was conducted to compare the efficacy of Ovsynch and kisspeptin protocol and need of fortification for treatment of postpartum anestrus. Fifty postpartum anestrus cows were selected. The animals were divided into four treatment groups (10 cows in each) and treated with Ovsynch (Group I), Ovsynch fortified with mineral mixture (MM) and bypass fat (BPF) (Group II), kisspeptin (Group III) and kisspeptin fortified with MM +BPF (Group IV). The remaining 10 cows served as control (Group V). The oestrus response was recorded on the basis of behavioral signs, physical signs and clinico-gynaecological changes. The blood was collected following treatment for hormonal estimation. The pregnancy was confirmed on day 45 post AI using ultrasound.

**Result:** The oestrus response was pronounced in Ovsynch as compared to kisspeptin groups. Further, fortification has enhanced the response. The serum oestrogen concentration increased following treatment and was significantly high on day of onset of oestrus with highest concentration in group II. The results indicate Ovsynch distinguishably succeeded in addressing postpartum anestrus with fortification enhances fertility response. However, kisspeptin also emerged as a potent candidate for the resumption of cyclicity.

**Key words:** Kisspeptin, Oestrogen, Ovsynch, Postpartum anestrus, Progesterone.

## INTRODUCTION

The enormous impact of postpartum anestrus in reproductive performance of an animal is well known. Factors such as reduced energy intake, poor body reserve and reproductive diseases can prolong the period of resumption of cyclicity (Peter *et al.*, 2009; Dixit *et al.*, 2020). The negative energy balance (NEB) results in poor release of reproductive hormones contributing in delayed postpartum oestrus and ovulation (Butler *et al.*, 2003). Such cows are in greater risk of being culled by dairy producers in order to upkeep the economic benefit (Esslemont *et al.*, 2001).

Attempts have been made to address prolonged postpartum anestrus in the dairy cattle using various protocol. Ovsynch program have been extensively evaluated for treatment of postpartum anestrus (Bhoraniya *et al.*, 2012; Dhami *et al.*, 2015). In recent years, new concepts for early onset of puberty, induction of oestrus using kisspeptin has been reported (Caraty *et al.*, 2007; Redmond *et al.*, 2011). Kisspeptin is a neuropeptide which is encoded by KiSS1 gene and its cognate. The KiSS1 gene is expressed in specific areas of the hypothalamus (Preoptic area and arcuate nucleus) that are critical for secretion of gonadal steroid hormone (Pielecka-Fortuna *et al.*, 2008). The kisspeptin stimulates pulsatile release of GnRH which in turn causes release of LH and FSH. Earlier study had demonstrated increased number of follicles at wave emergence, diameter of dominant follicle, oestrus response

<sup>1</sup>Department of Animal Reproduction, Gynaecology and Obstetrics, College of Veterinary Science, Khanapara, Guwahati-781 022, Assam, India.

<sup>2</sup>Department of Veterinary Physiology, College of Veterinary Science, Assam Agricultural University, Khanapara-781 022, Assam, India.

<sup>3</sup>Department of Veterinary Surgery and Radiology, College of Veterinary Science, Assam Agricultural University, Khanapara-781 022, Assam, India.

<sup>4</sup>Department of Animal Nutrition, Apollo college of Veterinary Medicine, Jaipur-302 031, Rajasthan, India.

<sup>5</sup>Department of Veterinary Pathology, College of Veterinary and Animal Sciences, Bihar Animal Sciences University, Arrabari, Kishanganj-855 107, Bihar, India.

**Corresponding Author:** M. Malik, Department of Veterinary Pathology, College of Veterinary and Animal Sciences, Bihar Animal Sciences University, Arrabari, Kishanganj-855 107, Bihar, India. Email: drmahmudamalik@gmail.com

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rate and duration of oestrus following kisspeptin administration. The other authors had advocated the use of

kisspeptin in anestrus animals (Pottapenjara *et al.*, 2018).

The reproductive physiology and secretion of reproductive hormones is directly associated with energy status of animals. It has been reported that the cows with low body fat reserves are unlikely to respond to hormonal therapy (Peter *et al.*, 2009). Earlier literature has reported beneficial effect of bypass fat (BPF) and mineral mixture (MM) on postpartum ovarian activity (Colazo *et al.*, 2009). Hence, could be useful in augmentation of hormonal protocol against postpartum anoestrus. Therefore, the present study was designed with aim to evaluate the efficacy of Ovsynch and kisspeptin protocol and need of fortification for the treatment of postpartum anestrus in crossbred cows. Further, to best of our knowledge, information in the regard of kisspeptin as a therapeutic measure in postpartum anestrus cows is scanty.

## MATERIALS AND METHODS

The study was approved by the Institutional Animal Ethics Committee of College of Veterinary Science, Assam Agricultural University, Khanapara, Assam, India.

### Experimental animals

The present study was conducted during the period of one year at Instructional Livestock Farm of College of Veterinary Science, Assam Agricultural University, Khanapara, Assam. Fifty postpartum anestrus crossbred cows were utilized for the study. The animals with the history of absence of cyclicity for more than 90 days after parturition were examined per rectum for ovarian activity. The cows having smooth ovaries with no palpable corpus luteum of any stage with quiescent genital organ were diagnosed as postpartum anestrus. The ultrasonography was performed in experimental animals to confirm the status of the animal as postpartum anestrus. The experimental cows were randomly divided into four treatment groups (group I, II, III and IV) comprising 10 cows in each. While, ten animals receiving no treatment served as control. All the cows were between 3 to 4 parity and subjected to similar feed and managemental conditions throughout the study.

### Treatment

#### Ovsynch protocol (group I)

The animals were subjected to Ovsynch protocol as described by Bhoraniya *et al.* (2012). The cows were injected with 0.020 mg of GnRH analogue (Buserelin Acetate, Gynarich®, Intas Pharmaceutical Ltd., Ahmedabad, India) on day 0 followed by 500 µg of cloprostenol sodium (Pragma™, Intas Pharmaceutical Ltd., Ahmedabad, India) on day 7 and 0.020 mg of GnRH on day 9 through intramuscular route. Fixed time AI was performed 16-24 hrs after 2<sup>nd</sup> GnRH injection.

#### Fortified ovsynch protocol (group II)

The cows were fed 50 g/cow/day MM (Agrimin® forte, Virbac Animal Health India Pvt Ltd., Maharashtra, India) and 150 g/cow/day BPF (Fatomax®, Intas Pharmaceutical Ltd.,

Ahmedabad, India) orally for a period of 21 days. The supplementation was followed by Ovsynch protocol.

#### Kisspeptin protocol (group III)

The animals were administered with kisspeptin-prostaglandin-kisspeptin protocol. The cows were injected with kisspeptin (Metastin 45-54 calbiochem cat: 445888 U.S.A.) @ 1.3 µg/kg body weight (Khan *et al.*, 2019) intravenously on Day 0 and followed by intramuscular injection of Cloprostenol sodium (Cloprostenol, Pragma™, Intas Pharmaceutical Ltd., Ahmedabad, India) 500 µg on Day 7 and again kisspeptin injection on Day 9 of treatment. Fixed time AI was performed 16-24 hrs after second kisspeptin injection.

#### Fortified kisspeptin protocol (group IV)

Animals were supplemented with MM + BPF followed by kisspeptin. The therapeutic protocol was followed as described earlier.

#### Control (group V)

The animals not receiving feed supplementation or treatment for induction of oestrus served as control.

### Assessment of oestrus response

All the experimental cows were closely observed during the period of treatment for the sign of oestrus. Oestrus was detected on the basis of behavioural, physical and clinico-gynaecological changes. The clinico-gynaecological examination was carried out 24 hrs after the PGF<sub>2α</sub> injection and then at every 12 hrs to confirm oestrus in the experimental groups. The ultrasonography was performed to confirm oestrus and to check the status of preovulatory follicle and corpus luteum. Duration of oestrus was calculated from the time of first to last appearance of estrus signs and recorded in hours (Lopez *et al.*, 2004).

### Endocrine profile

Blood samples were collected on day 0, 7, 9 and 10 of treatment from all the experimental animals. The serum was separated and stored at -20°C for serum oestrogen and progesterone assay. Serum oestrogen and progesterone level were analyzed using commercial oestradiol ELISA kit (Calbiotech ct., El Cajon, CA 92020, USA) and Progesterone ELISA Kit (Calbiotech ct., El Cajon, CA 92022, USA) as per manufacturer's instructions and the reading of optical densities were taken immediately after completion of assay procedure in microplate reader (Biorad, USA).

### Assessment of pregnancy rate

The pregnancy was determined 45 days post AI using real-time B mode ultrasound machine (M-SONOSITE, FUJIFILM inc Bothell, WA 98021-3904 USA).

### Statistical analysis

The data of behavioral, physical and clinico-gynaecological changes were compiled using Microsoft excel (version 10) and presented in percentage. The duration of oestrus and serum hormonal concentrations were presented as Mean±SEM using Microsoft excel (version 10). To estimate

the difference between treatment groups, the duration of oestrus and endocrine profile were analyzed using two-way ANOVA by GraphPad Prism statistical software. The Chi-square test was used to analyze the conception rate. The level of significant was considered as  $P < 0.05$ .

## RESULTS AND DISCUSSION

### Oestrus response

The oestrus response observed in the study was 70-80 per cent (Table 1). The oestrus response was observed in treated groups while, none of the cows in control group exhibited oestrus behaviour. The prominent sign of oestrus was observed in groups fortified with MM+BPF. Similar to the present results, Naikoo *et al.* (2016) and Sahoo *et al.* (2016) reported oestrus response as 66% (Ovsynch) and 71% (MM + BPF supplementation), respectively. The 83.33% oestrus response following administration of kisspeptin in buffalo was observed by Pottapenjara *et al.* (2018). However, no earlier work on effect of kisspeptin on oestrus response in cattle has been reported to our knowledge. The higher oestrus response observed in group II and IV might be due to nutritional supplementation prior to hormonal treatment. Minerals have been reported to have a beneficial role in endocrine system and play an important role in resumption of follicular growth (Kor *et al.*, 2013). The fortification might have increased the number of ovarian follicles and enhanced the follicular growth (Ambrose *et al.*, 2006; Colazo *et al.*, 2009). The oestrus response in group III is attributed to administration of kisspeptin. The kisspeptin is known as

potent secretagogue of GnRH which might be responsible for onset of cyclicity in anoestrus cows in the present study.

The duration of oestrus in the study ranged from 18.57 to 32.75 hrs. The duration of oestrus observed in group I and II was similar to earlier reports of Dudi *et al.* (2017) and Ratnaparkhi *et al.* (2020) using Ovsynch and fortified Ovsynch protocol, respectively. The prolonged duration of oestrus in group III and IV might be attributed to lower dose (1.3 µg/kg) of kisspeptin used in the present investigation. In contrast, lesser duration of oestrus (22.50 hrs) was observed following administration of Kisspeptin (20 µg/kg) by Pottapenjara *et al.* (2018) in buffaloes. The authors further elucidated that the kisspeptin induced LH release is short-term and lower than GnRH induced LH release. This might be the reason of prolonged oestrus duration in Kisspeptin group as compared to Ovsynch group in our study.

### Signs of oestrus

The various behavioural signs of oestrus observed in present investigation have been shown in Table 2. The number of cows showing bellowing, mounting, frequent urination, loss of appetite and restlessness behaviour were higher in group II. While, sniffing of vulva and raised tail were observed higher in group I and IV, respectively. The observed physical signs of oestrus in cows have been presented in Table 3. The population of cows with congested and swollen vulva was found higher in group II and IV. The copious quantity of mucus discharge was found greater in cows treated under group II. The clinico-gynaecological changes noted in animals responding to treatment with different oestrus

**Table 1:** Oestrus response and duration of oestrus in anestrus crossbred cows following treatment with different oestrus induction protocols.

| Groups    | No. of cows treated | No. of cows responded | Oestrus response (%) | Duration of oestrus (hrs) |
|-----------|---------------------|-----------------------|----------------------|---------------------------|
| Group I   | 10                  | 7                     | 70.00                | 18.57±1.02 <sup>a</sup>   |
| Group II  | 10                  | 8                     | 80.00                | 20.75±1.06 <sup>a</sup>   |
| Group III | 10                  | 8                     | 80.00                | 32.75±0.92 <sup>b</sup>   |
| Group IV  | 10                  | 8                     | 80.00                | 28.50±1.58 <sup>b</sup>   |
| Group V   | 10                  | 0                     | 0.00                 | -                         |

Data are presented as Mean±SEM. Mean bearing superscript a:b:c:d= $p < 0.05$  in a column differ significantly.

**Table 2:** Frequency of occurrence of various behavioural signs of oestrus in anestrus crossbred cows responding to different oestrus induction protocols.

| Behavioural signs     | Frequency of occurrence |             |             |             |
|-----------------------|-------------------------|-------------|-------------|-------------|
|                       | Group I                 | Group II    | Group III   | Group IV    |
| No. of cows responded | 7                       | 8           | 8           | 8           |
| Bellowing             | 1/7 (14.28)             | 4/8 (50.00) | 2/8 (25.00) | 3/8 (37.50) |
| Mounting              | 2/7 (28.57)             | 3/8 (37.50) | 1/8 (12.50) | 1/8 (12.50) |
| Sniffing of vulva     | 2/7 (28.57)             | 2/8 (25.00) | 1/8 (12.50) | 2/8 (25.00) |
| Frequent urination    | 3/7 (42.85)             | 5/8 (62.50) | 4/8 (50.00) | 5/8 (62.50) |
| Restlessness          | 3/7 (42.85)             | 5/8 (62.50) | 3/8 (37.50) | 4/8 (50.00) |
| Loss of appetite      | 1/7 (14.28)             | 2/8 (25.00) | 2/8 (25.00) | 1/8 (12.50) |
| Tail raising          | 1/7 (14.28)             | 1/8 (12.50) | 1/8 (12.50) | 2/8 (25.00) |

Figures in parentheses indicate percentage of animal exhibited signs of oestrus.

induction protocols have been reported in Table 4. On examination, the open cervix, uterine tonicity and presence of large follicles with soft consistency were found in all treated animals. However, the proportion of cows with good uterine tone and soft follicular consistency was greater in group II as compared to other groups.

The behavioural signs of oestrus were observed to be higher in groups supplemented with MM and BPF. The supplementation might have showed a beneficial effect on resumption of ovarian cyclicity in anestrus animals. It is well known that minerals serve as co-factors in the steroidogenesis and play an important role in resumption of ovarian activity (Smith and Akinbamijo, 2000). Similarly, feeding fat positively influence ovarian functions by enhancing the synthesis of steroidal hormones (Rahbar *et al.*, 2014). The better behavioural signs of oestrus following fortification might be due to large size of ovulatory follicle and higher oestradiol production. In the present study, fortification resulted in higher percentage of cattle showing positive clinico-gynaecological changes such as relaxed cervix, turgid uterus and presence of large follicles. The swollen vagina with congested mucosa was also observed better in groups supplemented with MM and BPF. Similarly,

copious discharge of cervico-vaginal mucus was found in the aforementioned groups. The fortification might have evoked better gonadal stimulation that resulted in higher elevation of oestrogen level which could bring about more conspicuous changes in the genitalia. The vaginal mucus discharge recorded (50-75%) were comparable to 61.7% observed by Negussie *et al.* (2002). In contrast, Nevkar *et al.* (2012) observed 45 per cent cows with mucus discharge, congested vulvar mucosa and swelling of vulva following treatment with Ovsynch protocol.

The changes in the genitalia were observed to be more pronounced in group I and II as compared to III and IV. The Ovsynch treatment in Group I and II might have evoked better gonadal stimulation leading to higher oestrogen level which could bring about more conspicuous changes in the genitalia (Roberts, 1971; Salisbury and Vandemark, 1961). As the oestrogen secreted from Graafian follicle is the reason of gradual relaxation of cervix and increased uterine tonicity (Jainudeen and Hafez, 1987). However, inferior results in Group III and IV treated animals might be due to low dose of kisspeptin (Chaikhun-Marcou *et al.*, 2019). Further, the clinico-gynaecological changes were more pronounced in cows supplemented with MM and BPF. The supplements

**Table 3:** Frequency of occurrence of various physical signs of oestrus in anestrus crossbred cows responding to different oestrus induction protocols.

|                              | Physical signs | Frequency of occurrence |              |             |              |
|------------------------------|----------------|-------------------------|--------------|-------------|--------------|
|                              |                | Group I                 | Group II     | Group III   | Group IV     |
| Vulva                        | Congested      | 5/7 (71.42)             | 6/8 (75.00)  | 5/8 (62.50) | 6/8 (75.00)  |
|                              | Pale           | 2/7 (28.57)             | 2/8 (25.00)  | 3/8 (37.50) | 2/8 (25.00)  |
|                              | Swollen        | 7/7 (100.00)            | 8/8 (100.00) | 7/8 (87.50) | 8/8 (100.00) |
|                              | Wrinkled       | 0/7 (0.00)              | 0/8 (0.00)   | 1/8 (12.50) | 0/8 (0.00)   |
| Quantity of vaginal mucus    | Copious        | 5/7 (71.42)             | 6/8 (75.00)  | 4/8 (50.00) | 5/8 (62.50)  |
|                              | Scanty         | 2/7 (28.57)             | 2/8 (25.00)  | 4/8 (50.00) | 3/8 (37.50)  |
|                              | Absent         | 0/7 (0.00)              | 0/8 (0.00)   | 0/8 (0.00)  | 0/8 (0.00)   |
| Consistency of vaginal mucus | Thin           | 5/7 (71.42)             | 6/8 (75.00)  | 3/8 (37.50) | 4/8 (50.00)  |
|                              | Watery         | 2/7 (28.57)             | 2/8 (25.00)  | 5/8 (62.50) | 4/8 (50.00)  |

Figures in parentheses indicate percentage of animal exhibited signs of oestrus.

**Table 4:** Clinico-gynaecological changes determined in anestrus crossbred cows responding to different oestrus induction protocol.

| Clinico-gynaecological changes |                        | Frequency of occurrence |              |              |              |
|--------------------------------|------------------------|-------------------------|--------------|--------------|--------------|
|                                |                        | Group I                 | Group II     | Group III    | Group IV     |
| Patency of cervix              | Open                   | 7/7 (100.00)            | 8/8 (100.00) | 8/8 (100.00) | 8/8 (100.00) |
|                                | Closed                 | 0/7 (0.00)              | 0/8 (0.00)   | 0/8 (0.00)   | 0/8 (0.00)   |
| Tone of uterus                 | Good                   | 3/7 (42.85)             | 5/8 (62.50)  | 2/8 (25.00)  | 4/8 (50.00)  |
|                                | Moderate               | 4/7 (57.14)             | 3/8 (37.50)  | 6/8 (75.00)  | 4/8 (50.00)  |
|                                | No tone                | 0/7 (0.00)              | 0/8 (0.00)   | 0/8 (0.00)   | 0/8 (0.00)   |
| Ovarian changes                | Large follicle (>9 mm) | 7/7 (100.00)            | 8/8 (100.00) | 8/8 (100.00) | 8/8 (100.00) |
|                                | Medium sized (5-8 mm)  | 2/7 (28.57)             | 3/8 (37.50)  | 5/8 (62.50)  | 5/8 (62.50)  |
|                                | Small (<5 mm)          | 4/7 (57.14)             | 5/8 (62.50)  | 8/8 (100.00) | 8/8 (100.00) |
| Consistency of follicular wall | Tense                  | 3/7 (42.85)             | 2/8 (25.00)  | 5/8 (62.50)  | 6/8 (75.00)  |
|                                | Soft                   | 4/7 (57.14)             | 6/8 (75.00)  | 3/8 (37.50)  | 2/8 (25.00)  |

Figures in parentheses indicate percentage of animals showing clinico-gynaecological changes.

might have positive influence on reproduction by altering both hormonal and gonal status (Mattos *et al.*, 2000; Smith and Akinbamijo, 2000).

### Oestrogen concentration

The serum oestrogen concentration at various days of treatment in different treatment groups has been presented in Table 5. The level of serum oestrogen varied significantly between treatment protocols on 7<sup>th</sup>, 9<sup>th</sup> and 10<sup>th</sup> with control group but remained similar ( $P>0.05$ ) at day 0. The serum oestrogen concentration increased significantly ( $P<0.05$ ) from day 0 to day 10 of treatment in all the treatment protocols. Except in group II and IV where serum oestrogen concentration was statistically non-significant on day 7 and 9. The level of oestrogen concentration was found higher in group II at day 7, 9 and 10. The oestrogen concentration was found statistically greater in group I and II as compared to group III and IV. However, the difference was non-significant ( $P>0.05$ ) between corresponding groups.

The gradual increase in mean serum oestrogen concentration was observed from day 0 to day 10 in all treated groups. This might be due to 1<sup>st</sup> GnRH or kisspeptin injection which might have caused emergence of new follicular wave pursuing into growth and development of dominant follicle. Highest level of serum oestrogen concentration recorded on day 10 in all treatment protocol might be due to presence of pre-ovulatory follicle. The concentration of oestrogen was highest in Ovsynch groups which are in conformity to oestrus response of the animals. Poor response in kisspeptin groups is again accounted to inappropriate dosage. The better steroid secretion and follicular development in fortified groups is indicated by higher oestrogen concentration in fortified groups as compared to non-fortified ones.

### Progesterone concentration

The serum progesterone concentration at 0, 7, 9 and 10 day of treatment in various groups has been presented in

Table 6. The serum progesterone level was found to be higher ( $P<0.05$ ) on day 7 in group I, II, IV while the difference was non-significant ( $P>0.05$ ) in group III and V. The progesterone concentration was statistically similar between various groups at day 0 and 9. The serum progesterone concentration on day 7 and 10 was higher ( $P<0.05$ ) in group II and IV, respectively. The level of progesterone was observed to be gradually increased from day 0 to day 7 of treatment and the progesterone level was found to be significantly higher ( $P<0.01$ ) in group I, II and IV. However, mean progesterone level was found not to differ significantly in different days of treatment in group III.

Increase in serum progesterone level observed in the present investigation upto day 7 might be due to formation of CL or luteinization of follicles present in the ovary following GnRH and Kisspeptin administration. The significant reduction in serum progesterone levels on day 9 and 10 might be due to parenteral administration of PGF<sub>2α</sub> on day 7 of treatment protocol leading to CL lysis which caused physiological declination of progesterone. However, no significant change in the level of progesterone was observed in different days of treatment in group III indicating absence of CL formation or luteinization of follicle due to low secretion of LH. In agreement to the present study, Chaikhun *et al.* (2019) reported that administration of kisspeptin at the dose rate of 1.3 µg/kg caused lower secretion of LH as compared to buserelin injection. Besides, kisspeptin at the dose rate of 20 µg/kg causes maximum elevation of LH (Pottapenjera *et al.*, 2018). Luteal activity as indicated by serum progesterone concentration on day 7 of treatment might be due lower dose of kisspeptin (1.3 µg/kg) which failed to cause ovulation or luteinization of follicles.

### Pregnancy rate

In the present investigation, the overall pregnancy rate was recorded highest in group II followed by group III and IV. However, lowest pregnancy rate was observed in group I (Table 7). The difference in number of cows those conceived

**Table 5:** Level of serum oestrogen (pg/ml) in crossbred cows responding to various oestrus induction protocols at different days of treatment.

| Groups    | DAY 0                   | DAY 7                    | DAY 9                    | DAY 10                   |
|-----------|-------------------------|--------------------------|--------------------------|--------------------------|
| Group I   | 39.87±0.81 <sup>D</sup> | 44.26±0.92 <sup>Ca</sup> | 46.97±0.64 <sup>Ba</sup> | 52.36±1.17 <sup>Aa</sup> |
| Group II  | 39.08±0.88 <sup>C</sup> | 45.50±0.56 <sup>Ba</sup> | 47.14±0.73 <sup>Ba</sup> | 53.46±0.20 <sup>Aa</sup> |
| Group III | 38.16±0.25 <sup>D</sup> | 40.67±0.54 <sup>Cb</sup> | 42.37±0.53 <sup>Bb</sup> | 44.63±0.55 <sup>Ab</sup> |
| Group IV  | 39.28±0.43 <sup>C</sup> | 41.79±0.68 <sup>Bb</sup> | 43.39±0.71 <sup>Bb</sup> | 45.29±0.33 <sup>Ab</sup> |
| Group V   | 38.17±0.33              | 38.24±1.05 <sup>c</sup>  | 38.30±0.42 <sup>c</sup>  | 38.22±0.40 <sup>c</sup>  |

Data are presented as Mean±SEM. Mean bearing superscript A:B:C:D= $p<0.05$  in a row and a:b:c:d= $p<0.05$  in a column differ significantly.

**Table 6:** Level of progesterone (ng/ml) in crossbred cows responding to various oestrus induction protocols at different days of treatment.

| Groups    | DAY 0                  | DAY 7                   | DAY 9                  | DAY 10                  |
|-----------|------------------------|-------------------------|------------------------|-------------------------|
| Group I   | 0.62±0.04 <sup>B</sup> | 2.92±0.62 <sup>Aa</sup> | 0.47±0.09 <sup>B</sup> | 0.40±0.13 <sup>Ba</sup> |
| Group II  | 0.67±0.06 <sup>B</sup> | 3.68±0.41 <sup>Aa</sup> | 0.58±0.13 <sup>B</sup> | 0.41±0.11 <sup>Ba</sup> |
| Group III | 0.66±0.07              | 0.77±0.03 <sup>b</sup>  | 0.63±0.13              | 0.61±0.06 <sup>ab</sup> |
| Group IV  | 0.69±0.04 <sup>B</sup> | 1.47±0.33 <sup>Ab</sup> | 0.70±0.05 <sup>B</sup> | 0.72±0.04 <sup>Bb</sup> |
| Group V   | 0.57±0.04              | 0.52±0.05 <sup>b</sup>  | 0.51±0.02              | 0.50±0.02 <sup>a</sup>  |

Data are presented as Mean±SEM. Mean bearing superscript A:B:C:D= $p<0.05$  in a row and a:b:c:d= $p<0.05$  in a column differ significantly



**Table 7:** Effect of different hormonal protocols on pregnancy rate in postpartum anoestrous cows.

| Groups    | No. of cows treated | No. of cows conceived | Induced estrus | Second cycle | Third cycle  | Overall pregnancy rate (%) |
|-----------|---------------------|-----------------------|----------------|--------------|--------------|----------------------------|
| Group I   | 10                  | 4                     | 28.57% (2/7)   | 40.00% (2/5) | 0.00% (0/3)  | 40                         |
| Group II  | 10                  | 6                     | 37.50% (3/8)   | 40.00%(2/5)  | 33.33% (1/3) | 60                         |
| Group III | 10                  | 5                     | 12.50% (1/8)   | 42.85% (3/7) | 25.00% (1/4) | 50                         |
| Group IV  | 10                  | 5                     | 12.50% (1/8)   | 57.14% (4/7) | 0.00% (0/3)  | 50                         |

Figures in parentheses indicate number of animals conceived.

did not differ significantly at induced estrus, second and third cycle. The pregnancy rate in cows inseminated at induced oestrus was higher in Ovsynch protocol. This might effectively persuade to generate LH surge by pre-ovulatory follicle. It was clearly mentioned that GnRH induced LH response is higher as compared to kisspeptin induced protocol (Pottapenjara *et al.*, 2018; Chaikhun-Marcou *et al.*, 2019). The lower dose of kisspeptin used in the study might have failed to generate LH surge (Decourt *et al.*, 2014; McGrath *et al.*, 2016). The higher pregnancy rate was observed in kisspeptin treated cows inseminated at second oestrus. The reason of better pregnancy rate at subsequent oestrus is still unclear. However, we could suspect that under given dosage the kisspeptin has successfully onset the cyclicity and has sustained hormonal milieu with fruitful conception in subsequent oestrus.

## CONCLUSION

The study utilized Ovsynch, fortified Ovsynch, kisspeptin and fortified kisspeptin protocol for the treatment of postpartum anoestrous cows. The Ovsynch resulted in higher overall pregnancy rate as compared to kisspeptin protocol. The behavioural, physical and clinico-gynaecological changes were much prominent in fortified groups; fertility results of this protocol varied. The kisspeptin with or without fortification led to prolonged oestrus and delayed ovulation in crossbred cows with dose rate of 1.3 µg/kg body weight. Further, the dosage failed to generate LH surge. However, in subsequent oestrus, higher number of animals conceived in kisspeptin group. The mechanism for this differential pregnancy rate is unclear and need to be explored. Despite of lower dose of kisspeptin showed higher efficacy to bring animal into cyclicity. Thus, kisspeptin protocol can appear as valuable tool in the advancement of the reproductive technology. However, the higher dose rate of kisspeptin is need to be targeted in future study.

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## REFERENCES

- Ambrose, D.J., Kastelic, J.P., Corbett, R., Pitney, P.A., Petit, H.V., Small, J.A. and Zalkovic, P. (2006). Lower pregnancy losses in lactating dairy cows fed a diet enriched in  $\alpha$ -linolenic acid. *Journal of Dairy Science*. 89(8): 3066-3074.
- Bhoraniya, H.L., Dhami, A.J., Naikoo, M., Parmar, B.C. and Sarvaiya, N.P. (2012). Effect of estrus synchronization protocols on plasma progesterone profile and fertility in postpartum anoestrous Kankrej cows. *Tropical Animal Health and Production*. 44: 1191-1197.
- Butler, W.R. (2003). Energy balance relationships with follicular development, ovulation and fertility in postpartum dairy cows. *Livestock Production Science*. 83(2-3): 211-218.
- Caraty, A., Smith, J.T., Lomet, D., Ben Said, S., Morrissey, A., Cognie, J., Doughton, B. and Clarke, I.J. (2007). Kisspeptin synchronizes preovulatory surges in cyclical ewes and causes ovulation in seasonally acyclic ewes. *Endocrinology*. 148(11): 5258-5267.
- Chaikhun-Marcou, T., Sothibandhu, P., Suthikrai, W., Jintana, R., Makoom, P., Suadsong, S. and De Rensis, F. (2019). Comparison of the effects of kisspeptin-10 or GnRH on luteinizing hormone secretion during the luteal phase of the oestrous cycle in swamp buffalo cows. *Buffalo Bulletin*. 38(1): 127-134.
- Colazo, M.G., Hayirli, A., Doepel, L. and Ambrose, D.J. (2009). Reproductive performance of dairy cows is influenced by prepartum feed restriction and dietary fatty acid source. *Journal of Dairy Science*. 92(6): 2562-2571.
- Decourt, C., Caraty, A., Briant, C., Guillaume, D., Lomet, D., Chesneau, D., Lardic, L., Duchamp, G., Reigner, F., Monget, P. and Dufourmy, L. (2014). Acute injection and chronic perfusion of kisspeptin elicit gonadotropins release but fail to trigger ovulation in the mare. *Biology of Reproduction*. 90(2): 36-41.
- Dhami, A.J., Nakrani, B.B., Hadiya, K.K., Patel, J.A. and Shah, R.G. (2015). Comparative efficacy of different estrus synchronization protocols on estrus induction response, fertility and plasma progesterone and biochemical profile in crossbred anoestrous cows. *Veterinary World*. 8(11): 1310-1316.
- Dixit, C.P., Bhuyan, D., Bhuyan, M., Ahmed, K., Haloi, S., Borpujari, D., Chakravarty, H., Ikpe, A.B. and Lyngdoh, M.N. (2020). Prevalence of various reproductive disorders in the foothills of Himalaya. *Journal of Animal Research*. 10(4): 635-640. DOI: 10.30954/2277-940X.04.2020.24.
- Dudi, V., Mehta, J.S., Chaudhary, A.K., Kumar, P., Kumar, A. and Ruhil, S. (2017). Efficacy of different estrus synchronizing protocols on estrus induction in postpartum lactating dairy cows. *Ruminant Science*. 6(2): 337-340.

- Esslemont, R.J., Kossabati, M.A. and Allcock, J. (2001). Economics of fertility in dairy cows. *Proceedings of Recordings and Evaluation of Fertility Traits in UK Dairy Cattle*. pp19-20.
- Jainudeen, M.R. and Hafez, E.S.E. (1987). *Reproductive Cycles: Reproduction in Farm Animals*. (5<sup>th</sup> ed.). Lea and Febiger, Philadelphia, USA.
- Khan, M.H., Perumal, P., Hazarika, S.B. and Ezung, E. (2019). Exogenous kisspeptin (kp-10) resumes cyclicity in postpartum anoestrus Mithun cows. *Indian Journal of Animal Science*. 89(8): 843-847.
- Kor, N.M., Khanghah, K.M. and Veisi, A. (2013). Follicular fluid concentrations of biochemical metabolites and trace minerals in relation to ovarian follicle size in dairy cows. *Annual Research and Review in Biology*. 3: 397-404.
- Lopez, H., Satter, L.D. and Wiltbank, M.C. (2004). Relationship between level of milk production and estrous behavior of lactating dairy cows. *Animal Reproduction Science*. 81(3-4): 209-223.
- Mattos, R., Staples, C.R. and Thatcher, W.W. (2000). Effects of dietary fatty acids on reproduction in ruminants. *Reviews of Reproduction*. 5(1): 38-45.
- McGrath, B.M., Scott, C.J., Wynn, P.C., Loy, J. and Norman, S.T. (2016). Kisspeptin stimulates LH secretion but not ovulation in mares during vernal transition. *Theriogenology*. 86(6): 1566-1572.
- Naikoo, M., Dhami, A.J. and Ramakrishnan, A. (2016). Effect of estrus synchronization on plasma progesterone profile and fertility response in postpartum suckled anoestrus Kankrej cows. *Indian Journal of Animal Research*. 50(4): 460-465. Doi: 10.18805/ijar.11164.
- Negussie, F., Kassa, T. and Tibbo, M. (2002). Behavioural and physical signs associated with oestrus and some aspects of reproductive performance in Fogera cows and heifers. *Tropical Animal Health and Production*. 34: 319-328.
- Nevkar, S.G., Amle, M.B., Birade, H.S., Gaikwad, S.M., Ulemale, A.H. and Yadav, M.M. (2012). Estrus Response and Fertility Rate in Induced Estrus Crossbred Cows Synchronized with Ovsynch Protocol. In: *National Symposium on Addressing Animal Reproductive Stresses through Biotechnological Tools*. pp 126, Nov 21-23, 2012. Assam, India.
- Peter, A.T., Vos, P.L.A.M. and Ambrose, D.J. (2009). Postpartum anoestrus in dairy cattle. *Theriogenology*. 71(9): 1333-1342.
- Pielecka-Fortuna, J., Chu, Z. and Moenter, S.M. (2008). Kisspeptin acts directly and indirectly to increase gonadotropin-releasing hormone neuron activity and its effects are modulated by estradiol. *Endocrinology*. 149(4): 1979-1986.
- Pottapenjara, V., Rajanala, S.R., Reddy, C., Gangineni, A., Avula, K., Bejjanki, S.K., Sathagopam, S., Kesharwani, S. and Velmurugan, S. (2018). Kisspeptin modulates luteinizing hormone release and ovarian follicular dynamics in prepubertal and adult Murrah buffaloes. *Frontiers in Veterinary Science*. 5: 149. doi: 10.3389/fvets.2018.00149.
- Rahbar, B., Safdar, A.H.A. and Kor, N.M. (2014). Mechanisms through which fat supplementation could enhance reproduction in farm animal. *European Journal of Experimental Biology*. 4(1): 340-348.
- Ratnaparkhi, A.R., Deshmukh, S.G., Birade, H.S., Kale, V.B., Harkal, S.B. and Jadhao, A.D. (2020). Comparative efficacy of synchronization protocols for improving fertility in postpartum crossbred dairy cows. *Haryana Vet*. 59: 23-26.
- Redmond, J.S., Macedo, G.G., Velez, I.C., Caraty, A., Williams, G.L. and Amstalden, M. (2011). Kisspeptin activates the hypothalamic-adenohypophyseal-gonadal axis in prepubertal ewe lambs. *Reproduction*. 141(4): 541-548.
- Roberts, S.J. (1971). *Veterinary Obstetrics and Genital Diseases (Theriogenology)*, (2<sup>nd</sup> ed.). CBS Publisher, New Delhi, India.
- Sahoo, J.K., Das, S.K., Sethy, K., Mishra, S.K., Swain, R.K., Mishra, P.C. and Satapathy, D. (2016). Effect of supplementation of mineral mixture and bypass fat on performance of crossbred cattle. *Journal of Animal Research*. 6(4): 611-618. DOI: 10.5958/2277-940X.2016.00071.1.
- Salisbury, G.W. and Vandemark, N.L. (1961). *Physiology of Reproduction and Artificial Insemination of Cattle*. W.H. Freeman, San Francisco, United States.
- Smith, O.B. and Akinbamijo, O.O. (2000). Micronutrients and reproduction in farm animals. *Animal Reproduction Science*. 60(1): 549-560.