RESEARCH ARTICLE

Bhartiya Krishi Anusandhan Patrika



Temporal Assessment of Crop Protection Agents in Water Bodies of Haryana Districts

S. Mishra¹, M.K. Singh¹, M.K. Saini¹, S. Alam¹, L.K. Thakur¹, I. Mukherjee¹, J. Kumar¹

10.18805/BKAP607

ABSTRACT

Background: The study was conducted to quantify the presence of crop protection agents in 233 water samples collected from water bodies situated in cities of Panipat, Karnal, Rohtak, Sonipat and Hisar districts of Haryana, India. Water samples were collected during different seasons the year for the period from 2014-2018.

Methods: The samples were processed by liquid- liquid partitioning method and quantified by Gas Chromatography and Mass Spectrometry. The water samples were screened for 55 pesticides.

Result: Estimation of the residues revealed that 23.5 and 6.6% of pesticides were detected in 68 and 165 monsoon and winter samples, respectively. Chlorpyrifos, pretilachlor, butachlor and lambda cyhalothrin were predominantly present in all the water samples analyzed. Chorpyrifos detected in water samples from Karnal were above standard limit prescribed by EPA 8081and8141 but some samples were below UAS Health advisory life time value.

Key words: Crop protection agents, GC-MS, Pesticide Residue, Temporal, Water bodies.

INTRODUCTION

Water is one of the most precious natural resources, which is essential for life. The presence of xenobiotics, namely pesticides in water is a matter of global concern. Several pesticides, belonging to different chemical class are being used in agriculture throughout the world Sneha *et al.* (2012). Pesticides move into water bodies due to wet precipitation Kumar *et al.* (2007), leaching, field runoff, municipal, agricultural and industrial disposal. In many developing countries water from rivers, canals, wells, tube wells are used both for drinking and irrigation purpose.

The contamination of the ground water with several of the organochlorine pesticides (OCPs) has been reported (Singh et al., 2005). Surface water contamination may impact eco-toxicological effect on aquatic flora and fauna and also on human health through food chain Miyamoto et al. (1990). Contamination of ground water resources by pesticides is a matter of grave concern Papadopoulou-Mourkidou et al. (2004). DDT, is under restricted use for disease vector control (UNEP 2003). The major issue arises in the countries where ground water is the source of drinking water for rural and adjacent urban areas (Tuxen et al., 2000).

The water supply in north of India is from the rivers, tributaries in the region. Haryana in northern part is mainly responsible for supply of water to the National Capital, Delhi. Haryana is predominantly an agricultural state in India extends between 27°39′N to 30°55′n latitude and 74°27′ to 77°36′E longitude and covers a total geographical area of 442100 ha. The state's economy is mainly based on agriculture and industry. The major sources of irrigation and drinking water of the region are Anangpur dam, Hathni Kund Barrage, Sutlej Yammuna link canal, Munak Canal, Agra Canal etc. The study highlights the contamination of pesticides in irrigation water bodies like wells, tube wells,

¹Institute of Pesticide Formulation Technology, Udyog Vihar, Gurugram-122 016, Haryana, India.

Corresponding Author: S. Alam, Institute of Pesticide Formulation Technology, Udyog Vihar, Gurugram-122 016, Haryana, India. Email: alam_samsul@rediffmail.com

How to cite this article: Mishra, S., Singh, M.K., Saini, M.K., Alam, S., Thakur, L.K., Mukherjee, I. and Kumar, J. (2023). Temporal Assessment of Crop Protection Agents in Water Bodies of Haryana Districts. Bhartiya Krishi Anusandhan Patrika. doi: 10.18805/BKAP607.

<u>Submitted: 14-11-2022</u> <u>Accepted: 28-08-2023</u> <u>Online: 04-10-2023</u>

canals, ponds and rivers of region from Panipat, Karnal, Rohtak, Sonipat and Hissar districts of Haryana, India.

MATERIALS AND METHODS

Geographical location

Total 233 water samples collected from the water bodies from cites of Panipat, Karnal, Rohtak, Sonipat and Hissar districts of Haryana, India. The water samples from irrigation system *i.e.* wells, tube wells, canal and river system were collected throughout the different seasons of the year for the period from 2014-2018. Fig 1 shows districts map of Haryana. Water samples were collected in one-Litre capacity pre-cleaned and rinsed amber-colored bottles. Samples were stored in refrigerator at a temperature of 4°C till analysis to prevent degradation of pesticide residues before processing.

Na₂SO₄, (AR-grade), sodium chloride (AR grade) dichloromethane, acetone, methanol and n-hexane solvents of pesticide residue grade purchased from Merck (Darmstadt, Germany). 1L separating funnel, 250 ml round bottom flask, volumetric flask of 100, 50, 10 ml was used

for standard stock solution and its mixture preparation. 1.5 ml sample vials were used for final sample extract injection to instrument. Turbo-Vap LV evaporator (Caliper Life Sciences, USA), Liquid dispensers (Coleparmer USA), adjustable pipette 1 to 5 ml (Eppendroff, USA) was used. An electronic weighing balance (Mettler Germany) with digital display was used to weigh the certified reference materials (CRM) and other reagents purchased from Sigma Aldrich, USA whose purity were above 99%.

The samples were extracted using conventional liquid-liquid partitioning method for the determination of organic pollutants in water [AOAC official methods (2019) and Veeraiah and Prasad (1996)]. The method was validated for its effectiveness. The LOD, LOQ recovery, repeatability was calculated. The samples were processed for residue estimation of different groups of pesticides *viz.* organochlorines (OC), synthetic pyrethroids (SP) and organophosphates (OP). The samples were extracted as per above method and were injected in gas chromatography–mass spectroscopy (GC-MS) at Institute of Pesticide Formulation Technology.

The water samples were screened for 55 pesticides (Table 1). The Certified Reference Material (CRM) were purchased from Sigma Aldrich (USA). Individual pesticide solution of 100 mg/L concentration was prepared in n-hexane. 55 individual pesticide standard stock solutions were prepared for the preparation of pesticides mixture of 10 mg/L. Working standards and were prepared by serially diluting 10 mg/L pesticides mixture stock as per the sensitivity of the instrument. The standard stock and working solutions were stored at 4°C.

Gas chromatography system (Agilent Technology 7890A) with DB-5MS fused silica capillary column (Agilent JandW, GC Column, 30 m \times 0.25 mm \times 0.25 µm) was used for quantitative analysis. Agilent Chem Station System software was used for instrument control and data analysis. Gases

were passed through purification filters which can trap oxygen and moisture, before supply to the column. Agilent mass spectrometer (5975C inert XL EI/CI MSD) was used for identification and quantification and confirmation of 55 pesticides. The mass spectrometer was operated in the positive ion electron impact energy of 70 eV and an emission current 60 μA. Full scan data were obtained with a mass range of m/z 45–500. Scanning interval and SIM sampling rate were kept at 0.5 and 0.2 s, respectively. The full scan TIC chromatogram of Standard Mixture (0.5 ppm) and Control water samples are given in Fig 2 and 3 respectively. The presence of pesticides residues were quantified and confirmed by comparing the retention time, peak area, m/z of the sample with those of the standards. The fragmentation patterns of detected pesticides are given in Table 2.

Liquid-liquid partitioning extraction followed by gas chromatographic detection (USEPA 1980; AOAC official methods (2019) and Veeraiah and Prasad (1996) was used for the determination of pesticide residues. The percent recovery was performed to evaluate the procedure, by spiking 55 pesticides mixture with known concentrations. For water samples, the mean recovery ranged from 75 to 90%, with a relative standard deviation between 5-20%, at 0.050 μ g/L level. The detection limit was 0.010 μ g/L. The efûcacy of the methodology was within the 95% conûdence level. The instrument is repatable with the precision (RSDr), precision (RSDwr) is below 5% and LOQ for the pesticides is 0.050 ppm (As per SANTE guideline).

RESULTS AND DISCUSSION

Total 233 water samples were analysed for the presence of 55 pesticides, which were collected from cities of Panipat, Karnal, Rohtak, Sonipat, Hissar and Narnaul districts of Haryana, India (Table 3). Samples were collected during

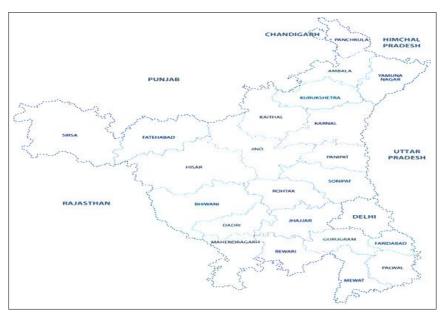


Fig 1: Geographical map of the state Haryana showing districts of water sampling.

Table 1: Continue...

(GV) µg/L 1 1 1 1 1 1 1 1 1 1 1 1 1	Pesticides	Pe	Pesticide detection range (µg/L)	tion range (µg/L)			EPA	МНО	USA	USA	USA Health	USA10-4
solutions BOL B	detected	Panipat	Karnal	Rohtak	Sonipat	Hissar	Narnaul	8081 and 8141	(GV) µg/L	(MCL) µg/L	(MCLG) µg/L	advisory life time µg/L	cancer risk µg/L
spot, s	Dichlorvos	BQL	BQL	BQL	BQL	BQL	BQL					ı	
Holy Boll Boll Boll Boll Boll Boll Boll B	Monocrotophos	BQL	BQL	BQL	BQL	BQL	BQL	1			ı	•	•
BOL BOL	Phorate	BQL	BQL	BQL	BQL	BQL	BQL	0.5			ı	•	•
Mathematical Mat	Dimethoate	BQL	BQL	BQL	BQL	BQL	BQL	0.5			ı	ı	•
Friently BOL	Diazinon	BQL	BQL	BQL	BQL	BQL	BQL	0.5		1	ı	9.0	
monition BQ1 BQ1 BQ1 BQ1 BQ1 BQ1 GQ1 GQ	Paraxon-methyl	BQL	BQL	BQL	BQL	BQL	BQL	1			ı	•	
rifos-methyl Boll Boll Boll Boll Boll Boll Boll Bo	Phosphomidon	BQL	BQL	BQL	BQL	BQL	BQL	1.0			ı	ı	
ny methyl BGL BGL BGL BGL BGL BGL BGL GGG GG	Chlorpyrifos-methyl	BQL	BQL	BQL	BQL	BQL	BQL	0.3			ı	•	
hion BQL BQL BQL BQL BQL BQL BQL BQL GG G G G G G G G G G G G G G G G G G	Parathion methyl	BQL	BQL	BQL	BQL	BQL	BQL	0.5			ı	ı	
put BQL BQL BQL BQL BQL BQL BQL QA C	Fenitrothion	BQL	BQL	BQL	BQL	BQL	BQL	0.5			ı	•	
rifes BGL 2-1515 BGL 80L 80L 10 0.3	Malathion	BQL	BQL	BQL	BQL	BQL	BQL	0.5			ı	100	•
n BQL	Chlorpyrifos	BQL	2-1515	BQL	30	BQL	_	0.3		1	ı	20	
with size and sequences and sequences are sequences and sequences and sequences are sequences as sequences are sequences as sequences are sequences as	Fenthion	BQL	BQL	BQL	BQL	BQL	BQL	0.5			ı	ı	•
hose BQL BQL BQL BQL BQL C.5 <td>Parathion</td> <td>BQL</td> <td>BQL</td> <td>BQL</td> <td>BQL</td> <td>BQL</td> <td>BQL</td> <td>0.5</td> <td></td> <td></td> <td>ı</td> <td>•</td> <td></td>	Parathion	BQL	BQL	BQL	BQL	BQL	BQL	0.5			ı	•	
hose Ball Ball Ball Ball Ball Ball Ball Bal	Chlorfenvinfos	BQL	BQL	BQL	BQL	BQL	BQL	0.5			•	1	•
fos BQL BQL BQL BQL BQL GO -	Quinolphos	BQL	BQL	BQL	BQL	BQL	BQL	1		1	•	ı	
fost BQL BQL BQL BQL TO <	Fenamiphos	BQL	BQL	BQL	BQL	BQL	BQL	0.5			ı	ı	•
8d BQL BQL BQL BQL BQL BQL BQL C	Profenofos	BQL	BQL	BQL	BQL	BQL	BQL	1.0			ı	ı	•
os BQL 122-439 BQL BQL BQL BQL BQL C	Ethion	BQL	BQL	BQL	BQL	BQL	BQL	0.5			ı	ı	•
os BQL BQL BQL BQL BQL BQL BQL BQL C	Trizophos	BQL	122-439	BQL	BQL	BQL	BQL	,			ı	ı	•
os BQL BQL BQL BQL BQL C <t< td=""><td>Edfinphos</td><td>BQL</td><td>BQL</td><td>BQL</td><td>BQL</td><td>BQL</td><td>BQL</td><td>1</td><td></td><td></td><td>ı</td><td>1</td><td></td></t<>	Edfinphos	BQL	BQL	BQL	BQL	BQL	BQL	1			ı	1	
osy BQL BQL BQL BQL BQL C <	Fipronil	BQL	BQL	BQL	BQL	BQL	BQL	1			ı	1	
zin BQL BQL BQL BQL BQL TS T	Anilophos	BQL	BQL	BQL	BQL	BQL	BQL	ı			i	1	•
OHE BQL BQL BQL BQL BQL BQL CH BQL	Buprofezin	BQL	BQL	BQL	BQL	BQL	BQL	1		•	ī	•	
CH BQL BQL BQL BQL BQL BQL BQL C-10 -	Phosalone	BQL	BQL	BQL	BQL	BQL	BQL	1.5		•	ī	•	
BQL BQL BQL BQL BQL BQL C-1 -	Alpha-HCH	BQL	BQL	BQL	BQL	BQL	BQL	0.05			ı	ı	•
ICH BQL BQL BQL BQL BQL BQL CO.5 -	Dicofol	BQL	BQL	BQL	BQL	BQL	BQL	0.10			ı	1	
a HCH (Lindane) BQL BQL BQL BQL BQL BQL C0.5 2 0.2	Beta-HCH	BQL	BQL	BQL	BQL	BQL	BQL	0.05			ı	1	
HCH BQL BQL <td>Gamma HCH (Lindane)</td> <td>BQL</td> <td>BQL</td> <td>BQL</td> <td>BQL</td> <td>BQL</td> <td>BQL</td> <td>0.05</td> <td>7</td> <td>0.2</td> <td>0.2</td> <td>0.2</td> <td>•</td>	Gamma HCH (Lindane)	BQL	BQL	BQL	BQL	BQL	BQL	0.05	7	0.2	0.2	0.2	•
or BQL BQL BQL BQL BQL BQL 0.03 0.03 0.04 00 - or BQL BQL BQL BQL BQL 0.50 20 2 0 - or BQL BQL BQL BQL BQL 0.55 2 3 200 BQL BQL BQL BQL BQL 0.04 0.03	Delta HCH	BQL	BQL	BQL	BQL	BQL	BQL	0.05			1	•	
or BQL BQL BQL BQL BQL 0.50 20 2 0 - ne BQL BQL BQL BQL BQL 0.5 2 3 200 BQL BQL BQL BQL BQL 0.04 0.03	Heptachlor	BQL	BQL	BQL	BQL	BQL	BQL	0.03	0.03	4.0	0	•	8.0
ne BQL BQL BQL BQL BQL 0.5 2 3 3 200 BQL BQL BQL BQL BQL 0.04 0.03	Alachlor	BQL	BQL	BQL	BQL	BQL	BQL	0.50	20	7	0	•	40
BQL BQL BQL BQL BQL 0.04 0.03	Atrazine	BQL	BQL	BQL	BQL	BQL	BQL	0.5	2	က	က	200	
	Aldrin	BQL	BQL	BQL	BQL	BQL	BQL	0.04	0.03		ı	1	0.2

:
<u>o</u>
2
ᆵ
Ŗ
_
믉
<u>a</u>
•

ı	•				0.2											•				•	
ı																					
ı	ı																				
Ī	ı	1	1	1	1	1	1		1	1	1	1	1	1	1		1	1	1		
1	20				0.03						ı	ı	ı	ı	ı		ı		ı		
	0.02	0.04	0.04		0.02	0.04	0.04		0.04	0.10	0.10	0.04		0.05	0.05	0.25	0.25	0.10	0.25	0.25	
BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	
BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	
BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	
BQL	BQL	BQL	BQL	2-279	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	09	BQL	BQL	BQL	
87-906	14	BQL	BQL	16-51	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	5-520	728-935	BQL	BQL	292	
BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	BQL	
Pretilachlor	Pendimethalin	O,P DDE	Alpha-Endosulphan	Butachlor	Dialdrin	P,P DDE	O,P DDD	P,P DDT	Beta- Endosulphan	P,P DDD	O,P DDT	Endosulphan Sulphate	Bifenthrin	Fenpropathrin	Lambda Cyhalothrin	Beta Cyfluthrin	Cypermethrin	Fenvalarate	Fluvalinate	Deltamethrin	

BQL- Below quantification limit- (<0.050 ig/L); GV- Guideline value; MCL- Maximum contaminant level; MCLG- Maximum contaminant level goal.

Table 2: Mass fragmentation of Pesticides detected in drinking water.

Pesticide detected	Molecular mass		Qualifier Ions	
1 colloide deleoted	Wolcodial Mass	Q1	Q2	Q3
Chlorpyrifos	350	97	197	199
Pendimethlin	281	252	162	281
Trizophos	313	172	161	97
Fipronil	437	369	367	351
Bifenthrin	181	166	141	-
Butachlor	312	160	176	57
Cypermethrin	209	163	181	91
Pretilachlor	312	162	238	176
Lambda cyhalothrin	450	181	197	208
Deltamethrin	253	181	77	93
Buprofezin	105	41	57	43

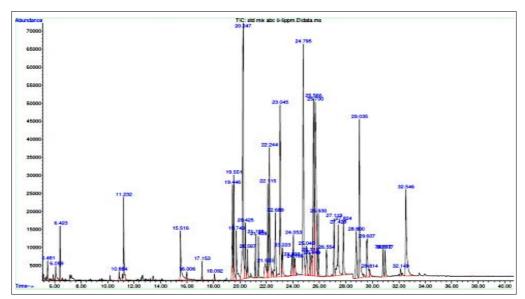


Fig 2: Standard mixture TIC chromatogram.

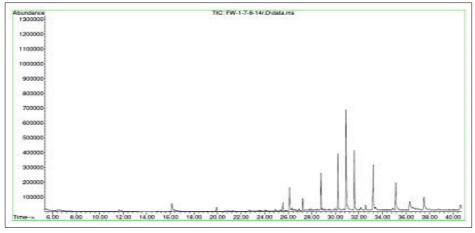


Fig 3: Control water sample TIC chromatogram.

pesticides Multi (%) 23.4 0.04 pesticide residues Total multi samples 7 α with multi pesticide No. of samples residues detected with % of samples pesticides 40.4 15.5 0.03 0.07 Contaminated samples 19 no. of samples detected in Pesticides Table 3: Summarized pesticide residues in water samples of northern India. samples no. of Total 70 43 45 30 26 15 samples collected No. of 10 10 တ Post monsoon Post monsoon Post monsoon Post monsoon Post monsoon Post winter Post winter Pre-winter Monsoon Monsoon Monsoon Season Winter Mar-2018 Month and Sep- 2017 April 2014 Feb-2016 Dec-2017 Jan-2016 Dec-2015 Sep-2017 Dec-2017 July-2018 Aug-2014 Dec-2014 Feb-2016 July 2014 Jan 2016 Jan-2016 Nov-2017 Oct-2018 Oct-2018 Jan-2016 Nov-2017 Jan-2016 Oct-2017 year Districts Panipat Narnaul Sonipat Rohtak Hissar Karnal

pre and post monsoon and pre and post winter season. Most of the crops are grown during these seasons due to conducive weather conditions. Intensive agriculture during this period leads to high irrigation and frequent use of fertilizers and pesticides. This may lead to runoff from fields into water bodies and aquifers. Temporal assessment of pesticides in water bodies during these months may give an insight into the levels of crop protection agents in water and their safety towards consumption for drinking purpose. The pesticides selected for analysis are based on the consumption pattern of pesticides in the area and the crops grown.

Analysis of results of Karnal districts indicate that 19 water samples showed the presence of pesticides, out of which multiple pesticide residues were detected in 11 samples. The pesticides detected in the samples were chlorpyrifos, triazofos, fipronil, bifenthrin, pendimethalin, butachlor, cypermethrin, pretilachlor, lambda-cyhalothrin, deltamethrin and buprofezin (Table 4). Chlorpyrifos was detected in 12, butachlor in 5, pretilachlor in 7 and lambda cyhalothrin in 7 samples. Water samples along the 230km str etch of the riverin Haryana were analyzed for the presence of organochlorine insecticide residues (Kaushik et al., 2010). Among 100 farmers, a survey was conducted in cauliflower and tomato cultivating areas of district Faridabad, Haryana, India from December 2012 to February 2013. Cypermethrin (62%) and profenofos (58%) were found as the most popular insecticides (Tyagi et al. 2015).

The range of contamination of pesticides in water samples was 2-1500 µg/L. Total 45 samples were collected from Rohtak in which 7 samples showed the presence of pesticides. Butachlor was detected in 7 samples (Table 4). From 30 and 15, samples from Sonipat and Narnaul, one sample each showed the presence of pesticides, (Table 3). A total 233 water samples were analyzed; out of which 70 samples were from Panipat, 43 samples from Karnal, 45 samples from Rohtak, 30 water samples from Sonipat, 26 samples from Hissar and 15 samples from Narnaul. Amongst the samples analyzed, 25 water samples showed the presence of at least one pesticide and in 11 samples multiple pesticide residues were recorded. Chlorpyrifos, butachlor, pretilachlor and cypermethrin were detected in 14, 12, 8 and 7 samples, respectively, whereas 2 samples showed the presence of triazofos and cypermethrin and 1 sample recorded the presence of five pesticides namely, fipronil, bifenthrin, pendimethalin, deltamethrin and buprofezin respectively (Table 3 and 5).

Temporal distribution of pesticides in water samples during different seasons is presented in Table 5. The pesticides detected during seasonal variation in 68 water samples of monsoon (pre and post) and 165 samples of winter (pre and post). It is observed that about 70% of crops are cultivated during winter season from mid-October to mid-April whereas only 30% of crops are grown during rainy season and localized farming is done during the months June to October. Thus, leading to increased inputs during winter season and subsequently prompting collection of

Table 4: Summary of pesticides detected and concentration range in water samples.

				o	-									
	Pesticides detect	Pesticides detected % of samples Samples with Conc	Samples with	Conc			Numbe	₃r of samples	detected:	Number of samples detected with pesticides				
Districts	in no. of	in no. of detected with multi pesticide range Chlorowrifos Bandimathlin Triazonhos Fibronii Bifanthrin Butachlor Cyparmathrin Pratilachlor	multi pesticide	range	Jornarifoe	Dendimethlin	Triazophos	Finconil Rife	anthrin B	utachlor Cyperm	ethrin Pretilachlo	ن	Deltamethrip Borofenzin	Anrofenzin
	samples	pesticides	residues	hg/L	5		50100	5				Cyhalothrii		5
Panipat	70	0												.
Karnal	47	16	1	1-1500	13	2	_	_	_	5	7	7	-	_
Rohtak	45	7	7	0.20-132						7	-		•	
Sonipat	30	-	,	BQL-30	_						,		ı	
Hissar	26	0	,								•		,	
Narnaul	15	-	,	BQL-1	_						,		ı	
Total districts	icts 233	25	13		4	2	-	-	_	12 2	80	7	-	-

season.
winter
and
monsoon
during
water
.⊑
residues
pesticides
with
detected
f samples
S
Details
5
able

		No. of	Total	No. of samples	No. of samples Pesticides detected % of samples	% of samples	No. of samples	Total no. samples	% of samples
Season	Districts	samples	no. of	no. of detected with	in total no. of detected with	detected with	with multi	with multi	with multi
		region wise	samples	pesticides	samples	pesticides	pesticide residues	pesticide residues pesticide residues pesticide residues	pesticide residues
Monsoon (Pre and post) Panipat	Panipat	15	89	ı	16	23.50		10	0.15
	Karnal	18		7			o		
	Rohtak	25		2			_		
	Sonipat	4							
	Hissar	9							
Winter (Pre and post)	Panipat	55	165		11	09.9		က	0.02
	Karnal	29		7			2		
	Rohtak	20		7			_		
	Sonipat	26		-			,		
	Hissar	20		ı			ı		
	Narnaul	15		-			,		

larger number of water samples in winter season than during the monsoon. A total of 68 samples were collected from Panipat, Rohtak, Karnal, Hissar and Sonipat during monsoon seasons (pre and post), among which 16 samples showed the presence of pesticides residues and 10 samples recorded the presence of more than one pesticide. Among the 165 samples of winter season, only 11 samples recorded pesticides and multiple pesticide residues were detected in 3 samples. The low pest incidence during the winter season leads to lesser use of pesticides and resulting in lower detection of pesticides in water samples during the winter months. In contrast, the pest and disease attack during the monsoon season is high due to the conducive climatic conditions, thus prompting high use of crop protection agents and leading to higher detection of pesticides. Regulatory limits by various regulatory bodies (Hamilton et al., 2003) has been compared with the contamination of the detected pesticides. The water samples of Panipat and Hissar region detected pesticide levels below the quantification level (Table 4 and 1). Chlorpyrifos was detected in the range of $1-1515 \mu g/L$, BQL (<0.050 $\mu g/L$) -30 $\mu g/L$ and BQL (<0.050 $\mu g/L$)-1 μg/L in Karnal, Sonipat and Narnaul samples, respectively. The level of concentration in 13 water samples of Karnal districts were above EPA 8081 and 8141 value of 0.3 µg/L but 4 samples were below UAS health advisory life time value of 20 µg/L. Chlorpyrifos detected in one samples from Sonipat was above EPA and USA health advisory life time, however one sample of Narnaul was above EPA but below USA Health advisory life time. Triazofos, pretilachlor, bifenthrin, fipronil and buprofezin were detected in the range of 122-439 μg/L; 87-906 μg/L, BQL (<0.050 μg/L-33μg/L; BQL ($<0.050 \mu g/L-18 \mu g/L$ and BQL ($<0.050 \mu g/L-2 \mu g/L$, respectively in Karnal water samples, no limit has been ascertained by any of the regulatory authorities. Butachlor pesticide detected in Karnal and Rohtak water samples ranged from 16-51 µg/L and 0.2-28 µg/L, respectively. Seven water samples from Karnal recorded the presence of lambda cyhalothrin in the range of 5-520 µg/L and were above the EPA value of 0.05 μg/L. Similarly, cypermethrin residues in 2 water samples of Karnal and 1 sample of Rohtak was in the range of 728-935 µg/L and 60 µg/L, respectively were above EPA value of 0.25 µg/L. In one sample from Karnal, deltamethrin was detected at 292 µg/L concentration which was above EPA value of 0.25 μg/L. A similar study conducted on groundwater samples from tubewells during the year 2002-2003 in Hissar revealed that synthetic pyrethroids like deltamethrin was detected in water samples varying from $0.017-0.06 \mu g/L$ and cypermethrin from $0.022-0.090 \mu g/L$. The most predominantly detected pesticide was chlorpyrifos, in the range 0.607-1.121 μg/L, (Kumari et al., 2008). Chlorpyrifos was detected from groundwater samples of the agricultural area of Amravati, Bhandara and Yavatmal region of Maharashtra in the concentration of 0.25, 0.11 and 0.18 µg/L, respectively (Lari et al., 2014). The studies conducted by Kumar et al. (2007), in Hissar district of Haryana revealed that the rain water were detected with cypermethrin (0.2-1.0 μ g/L), deltamethrin (0.1-0.8 μ g/L) and triazofos (0.05-0.009 μ g/L) pesticides.

CONCLUSION

Panipat, Karnal, Rohtak, Sonipat, Hissar and Narnaul are the prominent agricultural districts of Haryana, India. Sufficient numbers of samples were collected for analysis of pesticides from these districts of Haryana. Temporal and seasonal pesticide residue variation was focused in the study. Pesticide residues in the water samples from Panipat and Hissar regions were below the quantification levels. 40.4; 15.5; 0.03; and 0.07% of pesticides were present in Karnal, Rohtak, Sonipat and Narnaul samples, respectively. In 23.4 and 0.04% of water samples of Karnal and Rohtak multi pesticide residues were detected Temporal variation reveals that 23.5% and 6.6% of pesticides were recorded in 68 and 165 samples of monsoon and winter samples. 0.15 and 0.02% of water samples from Karnal and Rohtak area showed the presence of multiple pesticide residues Chlorpyrifos, pretilachlor, butachlor and lambda chalothrin were predominantly present in all water samples analyzed. Water samples from Karnal detected with chlorpyrifos were above EPA method 8081 B and 8141 but few samples were below UAS Health advisory life time. Some water samples from Karnal region, detected with chlorpyrifos, deltamethrin, lambda cyhalothrin were above EPA method 8081 B and 8141. One sample each from Sonipat and Narnaul showed the presence of chlorpyrifos above EPA method 8081Band8141. Cypermethrin residues in water sample from Karnal and Rohtak were above EPA method 8081Band8141 value.

ACKNOWLEDGEMENT

The authors are highly thankful to Directorate of Horticulture, Haryana and Director IPFT for their help and support.

Funding

This work was supported by Institute of Pesticide Formulation Technology.

Disclosure statement

No potential conflict of interest was reported by the author.

REFERENCES

- AOAC. (2019). Official Methods of Analysis of the Association of Official Analytical Chemists: Official Methods of Analysis of AOAC International. 21st Edition, AOAC, Washington DC.
- Hamilton, D.J., Ambrus, A., Dieterle, R.M., Felsot, A.S., Harris, U.A., Holland, P.T., Katayama, A., Kurihara, A., Linders, J., Unsworth, J. and Wong, S.S. (2003). Regulatory Limits for pesticide residues in water. Pure Applied Chemistry. 75(8): 1123-1155.

- Kaushik, A., Sharma H.R., Jain S., Dawra J., Kaushik C.P. (2010).
 Pesticide pollution of River Ghaggar in Haryana, India.
 Environment Monitoring Assessment. 160: 61-69.
- Kumar, B., Madan, V.K. and Kathpal, T.S. (2007). Pesticide residues in rain water from Hissar, India. Environmental Monitoring and Assessment. 133(1-3): 467-71.
- Kumari, B., Madan, V.K. and Kathpal, T.S. (2008). Status of insecticide contamination of soil and water in Haryana, India. Environmental Monitoring and Assessment. 136: 239-244.
- Lari, Z.S., Khan, N.A., Gandhi, K.N., Meshram, T.S. and Thacker, N.P. (2014). Comparison of pesticide residues in surface water and ground water of agriculture intensive areas. Journal of Environmental Health Science and Engineering. 12: 11. doi: 10.1186/2052-336X-12-11.
- Miyamoto, J., Mikami, N. and Takimoto, Y. (1990). The Fate of Pesticides in Aquatic Ecosystems. In: Environmental Fate of Pesticides. [Hutson, D.H., Roberts, T.R. (eds.)]. Chichester, England: Wiley. 123-147.
- Papadopoulou-Mourkidou, E., Karpouzas, D.G., Patsias, J., Kotopoulou, A., Milothridou, A., Kintzikoglou, K. *et al.* (2004). The potential of pesticides to contaminate the ground water resources of the Axios river basin in Macdonia, North Greece: Part 1. Monitoring study in the north part of the basin. Science of Total Environment. 321: 127-46.
- Singh, K.P., Malik, A., Mohan, D. and Takroo, R. (2005). Distribution of persistant organochlorine pesticide residues in Gomti River, India. Bulletin of Environmental Contamination and Toxicology. 74: 146-154.
- Sneha, D. and Bhimte, P.U. (2012). Meshram persistent organochlorine pesticide residues in ground and surface water of Nagpur and Bhandara district. Bionano Frontier. 5: 244-249.
- Tuxen, N., Tuchsen, P.L., Rugge, K., Albrechtren, H.J. and Bjerg, P.L. (2000). Fate of seven pesticides in an aerobic aquifer studied in column experiments. Chemosphere. 45: 1485-94. doi: 10.1016/s0045-6535(99)00533-0.
- Tyagi, H., Gautam, T., Prashar, P. (2015). Survey of pesticide use patterns and farmers' perceptions: A case study from cauliflower and tomato cultivating areas of district Faridabad, Haryana, India. International Journal Medipharm Research. 1(3): 139-146.
- USEPA (United States Environment Protection Agency). In: Analysis of Pesticide Residues in Human and Environmental Samples-A Compilation of Methods Selected for use in Pesticide Monitoring Programs; 1980. [Watts, R.R. (editor)]. EPA-600/80-038.
- UNEP, (2003). Global reports on regionally based assessment of persistent toxic substances, UNEP Chemicals, Geneva, Switzerland.
- Veeraiah, K. and Prasad, M.K.D. (1996). A study on the organochlorine pesticides residue input into Kolleru through Tammieleru river. Ecology Environmental and Conservation. 2: 83-86.