



Role of Molecular Markers in Crop Breeding: A Review

Rajitha Jayakumar Nair, Manoj Kumar Pandey

10.18805/ag.R-2322

ABSTRACT

Molecular markers are effective tools used to 'flag' the location of a specific gene or the inheritance of a definite trait. Markers are unique DNA fragments that can be identified inside the entire genome. The development of molecular markers combined with high throughput technologies have paved the way for achieving the desirable traits as well as induced biotic and abiotic stress tolerance in plant, which enhanced the crop breeding. Highly polymorphic molecular markers are developed for gene mapping, estimation of genetic diversity, finding out the evolution and phylogeny of crop, analysis of heterosis, assessment of diploid/haploid crops and genotyping of cultivars along with Marker Assisted Breeding (MAB)/Marker Assisted Selection (MAS). These are the most significant objectives for crop breeding. This review reveals about the role of various recently developed molecular markers in the improvement of crop. Molecular markers act as a "milestone" for the researchers who aim to enhance crop breeding.

Key words: Crop breeding, Evolution and phylogeny of crop, Genetic diversity, Gene mapping, MAS, Molecular markers.

Abbreviation: AFLP- Amplified Fragment Length Polymorphism, APCR- Arbitrarily Primed PCR, BSA- Bulk Segregant Analysis, CAPS- Cleaved Amplified Polymorphic Sequence, CDBP-CAAT box derived polymorphism, cDNA-AFLP- Complementary, DNA-amplified fragment length polymorphism, cM-Centimorgan, CMP- Coefficient of Marker Polymorphism, DArT- Diversity Array Technology, DH- Double Haploids, EST- Expressed Sequence Tag, GBS- Genotyping by Sequencing, GD- Genetic Distance, GZ- Genome Zipper, IRAP- Inter-retro transposon Amplified Polymorphism, ISSR- Inter simple sequence repeat, MAB/ MAS- Marker Assisted Breeding / Marker Assisted Selection, NGS- Next Generation Sequencing, PC- Protein Content, PCR- Polymerase Chain Reaction, PIC- Polymorphic information content, QTL-Quantitative trait loci, RAPD-Random Amplified Polymorphic DNA, RFLP- Restriction fragment length polymorphism, RILs- Recombinant Inbred Line, SCAR-Sequence Characterized Amplified Region, SCoT-Start codon targeted, SNP-Single Nucleotide Polymorphism, SSR-Simple sequence repeat, STMS-Sequence-tagged microsatellitemarkers,

Biotechnology is a branch of biology that manipulates living organisms or any part of particular living organism to develop animals, plants and microorganisms for any specific purpose. Since past two decades there has been a quick growth in the discipline of Plant Biotechnology and the different techniques used in it. Biotechnological methods produce an innovative, effective and profitable output or products. The techniques used in biotechnology decreases the use of chemicals in agriculture, increases productivity of food, decreases dismissive environmental effects of conventional methods and reduces the cost of unprocessed material. These subjects have implementation in the use of biological approach for the people as well as manage studies for proper knowledge of basic life activities.

Traditional method of Plant Breeding is an effective field of life sciences. This method depends on selection of genotypes and genetic variation which are used to develop the attributes demanded or needed by the farmers and the people. Introduction of new resistance genes for abiotic and biotic resistance from different sources like related plant varieties or gene bank has also developed the traits of crops. However, conventional breeding techniques have been fortunate in producing improved genotypes of different crops, but the new advancement in molecular breeding enhances the development of genotypes in a very less duration of time. The field of plant breeding can be improved by new technologies involved in molecular breeding or Plant

Department of Genetics and Plant Breeding, School of Agriculture, Lovely Professional University, Phagwara-144 411, Punjab, India.

Corresponding Author: Manoj Kumar Pandey, Department of Genetics and Plant Breeding, School of Agriculture, Lovely Professional University, Phagwara-144 411, Punjab, India.
Email: manoj.22848@lpu.co.in

How to cite this article: Nair, R.J. and Pandey, M.K. (2021). Role of Molecular Markers in Crop Breeding: A Review. *Agricultural Reviews*. DOI: 10.18805/ag.R-2322.

Submitted: 06-07-2021 **Accepted:** 21-09-2021 **Online:** 08-10-2021

Biotechnology. The genetic composition of organisms was easily understood by the discoveries of Polymerase Chain Reaction (PCR) by Kary Mullis in 1983 (Mullis, 1990) and restriction enzyme by Smith and Wilcox (Smith and Wilcox, 1970). These discoveries achieved presumed genetic fingerprint. These experiments are done by the dissociation of DNA fragments on a gel electrophoresis that is obtained through a selective amplification of DNA utilizing Polymerase Chain Reaction (PCR) and by the segregation of DNA with enzymes.

Molecular markers

Molecular markers comprise of certain molecules, which manifest simply detectable dissimilarities between various

strains of a species or between different species. A molecular marker is described as a DNA fragment or gene with a familiar site on a chromosome and correlated with a specific gene or character.

Molecular markers are basically classified into two groups, viz, based on hybridization and based on Polymerase Chain Reaction (PCR). Types of molecular markers based on hybridization are: RFLP, microsatellite and minisatellite. Various molecular markers based on PCR are: Arbitrarily Primed PCR (AP-PCR), Start Codon Targeted Marker (SCoT), Random Amplified Polymorphic DNA (RAPD), Inter simple sequence repeat (ISSR), Single nucleotide polymorphism (SNP), Simple sequence repeat (SSR), Amplified Fragment Length Polymorphism (AFLP), Cleaved Amplified Polymorphic Sequence (CAPS) and Expressed Sequence Tag (EST). And markers based on DNA sequencing like Single Nucleotide Polymorphism (SNP) marker. These markers are categorized based on their expressions; they are: -

- A. Dominant markers: dominant markers are designated as dominant markers if one form of the character, which is to be marked is correlated with the marker whereas another form of the character is not correlated with any marker. Dominant markers do not differentiate between homozygote and heterozygote, for e.g.: RAPD
- B. Co-dominant markers: are called as co-dominant markers if both form of the characters, which are to be marked are correlated with the marker. Co- dominant marker can differentiate between homozygote and heterozygote. For e.g.: RFLP, SSR, SNP, EST, etc.

Important uses of molecular markers in Crop Improvement:

Gene mapping

The development of huge number of new molecular markers provided a deep insight about the construction of high-resolution map and molecular breeding procedures in plants (Singh *et al.*, 2017, Griffiths *et al.*, 2000, Mishra and Tomar.,

2014). Molecular markers are highly used for construction of genetic linkage mapping. It is also used in recognition of useful alleles in the variety or wild species. A molecular marker acts as a site of heterozygosity for some set of silent DNA differentiation, which are not associated with any quantifiable phenotypic variation. Such a locus of DNA, when in heterozygous form, can be utilized in gene mapping analysis like a conventional pair of heterozygous alleles can be used (Griffiths *et al.*, 2000). Hence, the use of molecular markers, such as SSR, SNP, DArT, Microsatellite, ISSR, RFLP, RAPD, etc. has facilitated mapping of gene, QTLs, gene discovery and various other advancements in plant breeding. Mapping of quantitative trait loci QTLs for grain protein content (PC) in rice useful for biofortification was studied using 98 molecular markers. Seven markers out of ninety-eight markers were strongly linked QTLs for grain protein content in rice (Pradhan *et al.*, 2019). Hybrid necrosis genes, namely, Ne1 and Ne2 were mapped through microsatellite markers in wheat. Genetic analysis showed that Alsen variety had ne1ne1Ne2Ne2 genotype and SHW had Ne1Ne1 ne2ne2 genotype. Xbarc74 and Xbarc55, microsatellite markers were used to map the genes in backcross populations. (Chu *et al.*, 2006). High resolution mapping of Barley Leaf Rust (*Puccinia hordei*) resistance gene, *Rph_{MBR1012}* was studied using 56 molecular markers (Fazlikhani *et al.*, 2019). Quantitative trait loci (QTL) identification was performed across different environment for yield and its component and fibre quality characters in cotton (Ramesh *et al.*, 2019; Diouf *et al.*, 2018; Shang *et al.*, 2016). CottonSNP63k Illumina infinium SNP array is widely used for genotyping different RILs and the parents (Ramesh *et al.*, 2019; Li *et al.*, 2016). Phenotypic variance from five different environments were analysed using composite interval mapping which showed a total fifty-six QTL justifying phenotypic variances in the range of 8.18%-28.91% and 34 QTL out of 64 QTL resulted in QTL X Environment interactions, which were identified through multi-environment trial analysis (Ramesh *et al.*, 2019). QTL X Environment

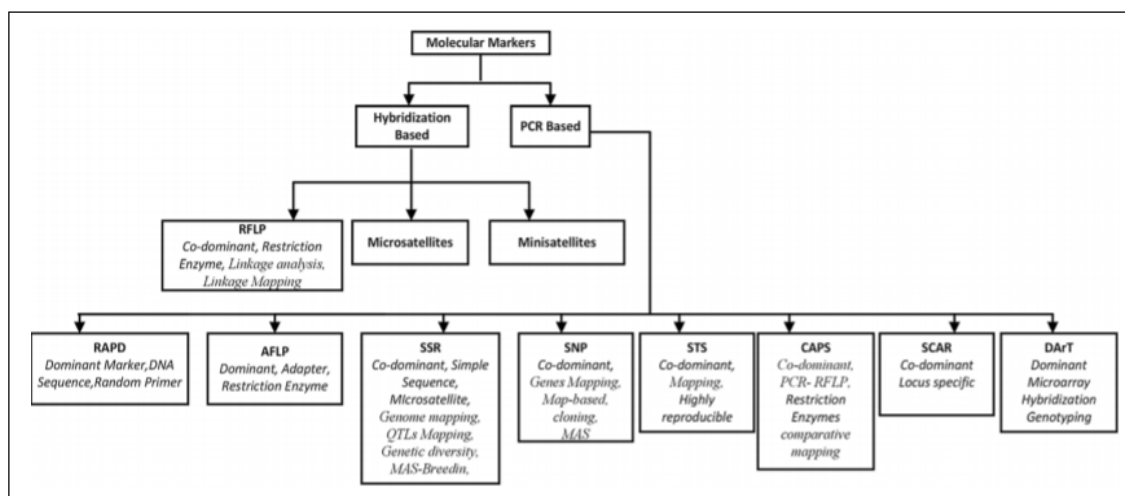


Fig 1: Types of molecular markers used in crop breeding (Gouda *et al.*, 2020).

interactions are detected for fibre quality through composite interval mapping which identified 62 new stable QTL across different environments (Shang *et al.*, 2016). Yield traits and morphological features were identified using molecular markers in maize. 346 SNPs and 623 SilicoDART (969 markers) out of 49,911 identified polymorphic molecular markers were associated with yield traits and analysed morphological traits (Tomkowiak *et al.*, 2019). DARTseq is a combination of Next Generation Sequencing (NGS) and DART (diversity array technology) complexity reduction methods (Tomkowiak *et al.*, 2019, Kilian *et al.*, 2012, Cruz *et al.*, 2013, Sansaloni *et al.*, 2011). The DARTseq method is used between others to identify SNPs and gives a large population of so-called silicoDARTs. SilicoDARTs have a dominant trait as the variability is found by the single point mutation (Sansaloni *et al.*, 2011). Hence, silicoDART and SNP markers can be used to group lines with incomplete data of origin and group lines in terms of origin of a species as well as in determining the applications in the selection of

parents for crossing in heterosis and for predicting the hybrid formula in maize (Tomkowiak *et al.*, 2019). An ameliorated genetic map with 1205 loci, having an average space of 2.2cM between loci and spanning 2598.3cM was developed in the recombinant inbred line (RILs) (TAG 24 x ICGV 86031) varieties of groundnut using a marker of high-density 58K SNP "Axiom_Arachis" array. Analysis of QTL was performed from the phenotypic data made for twenty drought tolerance and two iron deficiency tolerance related parameters at two locations in India and from 8 seasons, which resulted in nineteen major main- effect QTLs 10.0 to 33.9% phenotypic variation for iron deficiency tolerance and drought tolerance related parameters (Pandey *et al.*, 2021).

Estimation of genetic diversity

Recent progress in the technology of molecular markers act as an important tool for profundity of genetic diversity and increased the knowledge about breeding strategies (Ramesh *et al.*, 2020). Markers are used for estimation or

Table 2: Differentiation of some of the commonly used markers are as follows:

Molecular Markers	Type (Dominant / Co-dominant)	Amplification of marker/ technique used for identification	References
Restriction fragment length polymorphism (RFLP)	Co-dominant	Depends on point mutation in a restriction site	Botstein <i>et al.</i> (1980)
Random amplified polymorphic DNA (RAPD)	Dominant	Point mutation at primer annealing site in the specific region of a DNA strand	Williams <i>et al.</i> , 1990, Welsh and McClelland, 1990
Sequence Tagged Sites (STS)	Co-dominant	Depends on mutation at primer annealing site in the specific region of a DNA strand.	Olson <i>et al.</i> , 1989
Sequence characterized amplified region (SCAR)	Co-dominant	Depends on mutation at primer annealing site in the specific region of a DNA strand.	Paran and Michelmore, 1993
Amplified fragment length polymorphism (AFLP)	Dominant	Depends on mutation at primer annealing site in the target DNA and change in restriction site in the target DNA	Vos <i>et al.</i> , 1995
Cleaved amplified polymorphic sequence (CAPS)	Co-dominant	Depends on: -1. Mutation at primer annealing site in the target DNA.2. Change in restriction site in the target DNA.	Konieczny and Ausubel., 1993
Simple Sequences Repeats (SSRs)/ microsatellites	Co-dominant	Differences in the number of repeats of motif	Litt and Luty, 1989, Akkaya <i>et al.</i> 1992 in plants
Diversity Arrays Technology (DART)	Dominant	Microarray hybridization, DART arrays are produced from genomic libraries through amplification of candidate or random clones	Wenzl <i>et al.</i> , 2004
Single nucleotide polymorphism (SNP)	Co-dominant	Point mutation along with sequence information	Rafalski. 2002

assessment of genetic diversity in germplasm collection, advanced breeding material and other cultivars which helps in the characterization of germplasm, developing PGR information system and for evolving varietal information structure. SSR, SCoT, CBDP, DArT, ISSR, SNP, CAPS, SCAR, RFLP, AFLP, RAPD, etc. are some of the different markers, which are extensively used for the estimation of crop genetic diversity (Hossain *et al.*, 2020, Gaballah *et al.*, 2021, Ghobadi *et al.*, 2021, Kasoma *et al.*, 2020).

Genetic diversity was estimated for 16 rice varieties under water stress condition using SSR markers for drought tolerance. The varieties Giza 178, Giza179 and GZ1368-S-5-4 were found to be drought tolerant (Gaballah *et al.*, 2021). Start codon targeted (SCoT) polymorphisms and CBDP CAAT box-derived polymorphism (CBDP) are effective markers for the estimation of genetic diversity in hexaploid wheat. (Ghobadi *et al.*, 2021). Genetic diversity was successfully estimated in 48 accessions of barley (introduced from ICARDA, Lebanon) using 150 SSR markers, out of which 51 SSR marker showed polymorphism that were utilized for final analysis. These 51 SSR markers generated 158 polymorphic loci in the form of genotypic data with a mean of 3.275 alleles per SSR locus. Hence, the existence of favourable allelic diversity was confirmed which is important for estimation of genetic diversity (Kumar *et al.*, 2020). DArT sequencing-derived single nucleotide polymorphism markers revealed the genetic diversity of 59 genotype of maize for economic characters and resistance to the fall armyworm pest. A moderate level of genetic diversity among the genotypes were generated when evaluated using SNP markers as the mean gene diversity were 0.29 and Polymorphic information content (PIC) were 0.23. ZM 7114, ZM 4236 and Pool 16 were the maize genotypes with favourable agronomic traits and fall armyworm resistant (Kasoma *et al.*, 2020). Thus, recently developed microarray-based molecular marker technology shows high efficiency in determining the genetic diversity in maize (Kasoma *et al.*, 2020, Badu-Apraku *et al.*, 2021, Zebire, 2020). A large genetic variation in data set using

CottonSNP63K array provided an opportunity to differentiate *G. hirsutum* from other species of *Gossypium* and even distinctly separated wild types from cultivated types of *G. hirsutum*, and additionally, SNPmarker identified the loci associated with seed nutritional characters (Hinze *et al.*, 2017). Thirteen SSR markers were used to estimate genetic diversity. 119 genotypes were studied across two locations to evaluate agronomic characters and leaf spot and rust disease susceptibility. Moderate level of genetic diversity with 0.34 and 0.63 mean polymorphic information content and gene diversity, respectively which proved genotypes ICG 12725, ICGV-SM 16608, ICGV-SM 16575 and ICGV-SM 06737 are useful for the improvement of groundnut (Daudi *et al.*, 2021).

Evolution and phylogeny of crop

Advancement in the technologies of molecular markers have enhanced facts related to the genetic structure of a crop. Earlier, the evolution was studied on the basis of morphological or geographical changes between the crops, but, nowadays molecular markers are largely used to reconstruct a genetic map to reveal information about phylogeny and evolution of a crop (Nadeem *et al.*, 2018). Chloroplast markers are considered to be ideal for the evaluation of plant phylogeny, because of their stable and simple genetic construct (Dong *et al.*, 2012). Phylogenetic analysis is the organization of various species in a group based on their genetic relationship, which indicates the degree of genetic variation. Pattern of evolution of various species can be from this groups using molecular markers (Singh and Singh, 2015). A map of genome variation in rice proved that *Oryza sativa* L. (Japonica rice) to be the first domesticated rice species from a particular population of *O. rufipogon* and a cross made between *japonica rice* and wild local rice eventually developed the *Oryza sativa* (Indica rice) (Oka 1988, Cheng *et al.*, 2003, Huang *et al.*, 2012). Phylogenetic study revealed that SNPs are fixed by ancestors of *indica-japonica*, which resulted that 45% of the ancestral alleles of SNPs are fixed in *japonica* and 55% in

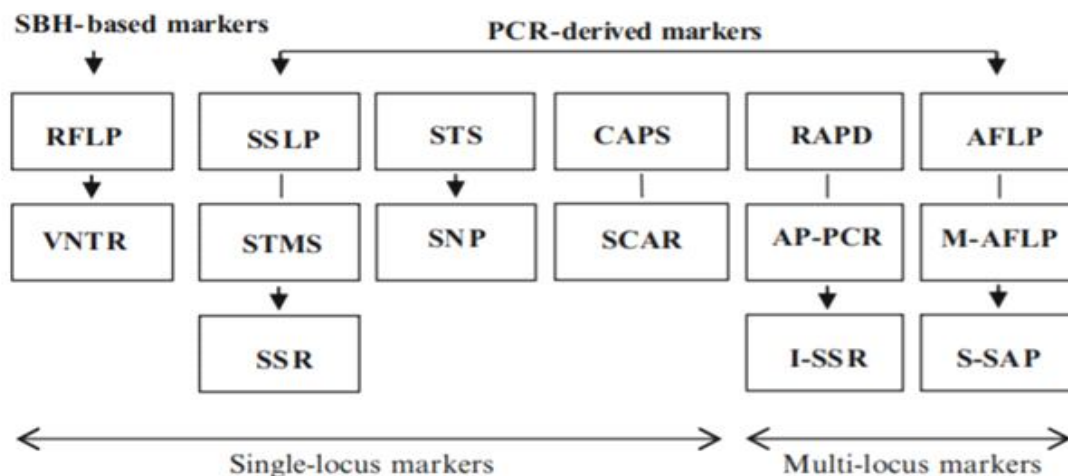


Fig 2: categorization of highly used molecular markers (Barcaccia, 2010).

indica (Huang *et al.*, 2012). Zhang *et al.*, 2021 studied genome distribution patterns in *Triticum aestivum* and its diploid and tetraploid progenitors comparing the numbers of coding sequences *i.e.*, AAC, AAG, AGC and AG. SSRs sequence evolution carried out for the identification of different chromosomes which in result indicated more sensitivity of B genome expansion and elimination during wheat evolution. Phylogenetic study of 14 cereals (4 wheat, 3 barley, 3 rice, 2 maize and 2 sorghum) using 10 ISSR-PCR marker, 15 RAPD-PCR marker and NTSYS-pc program data of both sets evinced wheat is more closely related to barley than rice followed by maize and sorghum (Rabey *et al.*, 2015)

Analysis of heterosis using molecular markers:

Heterosis describes the superiority of F_1 (progeny) over both its parents. The use of polymorphic molecular markers for the estimation of heterosis have gradually increased over last few decades. Molecular markers are effectively used in the analysis of heterosis in various population structure (Gonzalez *et al.*, 1999)

Heterosis was successfully predicted in 40 F_1 rice hybrids by using 25 EST- SSR and morphological markers Heterosis was analyzed based on correlation between standard heterosis and coefficient of marker polymorphism (CMP), in which CMP value ranged from 0.40-0.80 (Pavani *et al.*, 2018). Nie *et al.*, 2019 concluded that SNP (90K SNP chip) is an effective marker to accurately predict heterosis based on genetic distance (GD) and perfectly allocates the parents to heterotic groups in wheat. The experiment performed by Geng *et al.*, 2021, also revealed that SSR and SNP marker serves as an effective tool in predicting the heterosis based on parental GD as well as assigns the varieties into heterotic groups and even provides knowledge for selection of parents in hybrid breeding of cotton. A study was conducted to analyze the heterotic group and patterning in quality protein maize (QPM) in 3 inbreds of maize using SSR markers clustered the better hybrids together but in another sub clusters and flint X dent was the superior heterotic pattern (Rajendran *et al.*, 2014). Heterosis in barley was assessed through cDNA- AFLP marker in 48 crosses. 5 TDFs revealed the significance of heterosis of 1000- kernel weight (Zhang *et al.*, 2015).

Assessment of diploid/haploid crops and genotyping of cultivars

Haploids are described as crops having single set of chromosomes, diploids are crops having two copies of homologous chromosomes each and double haploids (DH) are crops obtained from a single pollen grain and artificially doubled to produce homozygous diploids. This DH or haploid crops are utilized as a mapping population for QTL mapping and other genetic studies (Khush and Virmani, 1996). Radi *et al.*, 2020 used an integrated protocol for the identification of haploids in a hybrid population of early seedling stage by using *R1-navajo* (*R1-nj*) phenotypic marker or anthocyanin

color marker (Chaikam *et al.*, 2015, Melchinger *et al.*, 2013) associated with genome size determination and SSR marker were used at pre-seedling stage for the assessment of haploid progenies in maize inbred lines. Total 38 STMS (Sequence-tagged microsatellite) markers identified heterozygotes from 200 DHs, which were derived from 'BS6444G', an elite indica rice hybrid. Out of 200 DHs, 9 DH line yielded higher than hybrid parents (Naik *et al.*, 2017). Inter-retro transposon amplified polymorphism (IRAP) markers proved to be efficient tools for fast genetic fingerprinting and screening of large germplasm of cotton. The marker works efficiently in identifying the diploids from tetraploid cotton as well as differentiated the cultivars. So as to achieve the data on genetic diversity, 17 diploid and tetraploid *Gossypium* accessions were used for the genetic fingerprinting through the manipulation of IRAP markers, which resulted in moderate (0.0–18%) to high (45–80%) genetic variability (Noor mohammadi *et al.*, 2018).

Marker Assisted Breeding (MAB)/ Marker Assisted Selection (MAS)

MAS/MAB is the indirect selection of selected or desired plant phenotype depending on the closely linked DNA marker. MAS/MAB is an efficient molecular tool for breeding, in which markers linked with the desired genes are used for indirect selection for that gene in non-segregating or segregating populations. MAS is an important method for the selection of traits that are difficult, like, biotic and abiotic stress tolerance in a crop (Datta *et al.*, 2011, Das *et al.*, 2017). Marker assisted selection successfully combines tolerance/resistance to various biotic and abiotic stresses as well as maintains grain quality and higher yield in rice. The experiment was conducted utilizing genes conferring resistance against bacterial leaf blight (Xa4, xa5, xa13, Xa21), blast (Pi9), gall midge (Gm4, Gm8) and drought tolerance QTLs (qDTY1.1 and qDTY3.1). 7 introgression lines (ILs) of rice having 7-10 QTLs were developed using MAS in the context of swarna with QTLs of drought. Out of these, 3 ILs *viz.* IL6 and IL7 were resistant/tolerant for multiple abiotic and biotic stresses in the field and glasshouse conditions. MAS methods were used to develop purple-grained cultivars of bread wheat. The breeding scheme initiated from crossing of lines and elite cultivars (recipient) with donor lines which had the complementary genes Pp-D1 and Pp3. Pp genes controls the anthocyanin pigments which results in purple colored grain (Gordeeva *et al.*, 2020). Lei *et al.*, 2020 newly developed a PCR based co-dominant marker identified the 17-kb deletion of *Hordeum vulgare* ethylene response factor (ERF) gene, that is seen on 7HL chromosome on the *nud* locus which is responsible for naked barley. SNP markers can be successfully used in the MAS of upland cotton because a *SNP_GH1570* was found totally segregated from fruiting branches character and SSR marker BNL3232 was linked to the short fruiting branches was screened by BSA method (Zhang *et al.*, 2018). MAB method was used to pyramid the β -Carotene (*crtRB1*)

and Opaque-2 (O2) genes in maize. 236 SSR markers equally spread across the maize genome were used for the background selection. *CrtRB1 32 TE* and *umc1066* markers were used for *crtRB1* and O2 gene, respectively for the foreground selection. The results showed increase in tryptophan (0.073-0.081%), lysine (0.294-0.332%) and β -carotene (6.12-7.38 $\mu\text{g/g}$) content (Chandran *et al.*, 2019).

CONCLUSION

Molecular marker plays an important role in plant breeding or crop improvement. A desirable molecular marker should have high polymorphism, frequent occurrence, should be easy to use and should be quick, co-dominant inheritance, equally dispersed all over the genome, high transferability and reproducibility, less expensive and phenotypically neutral. The last few years have revolutionized the molecular marker technology from RAPD to DArT markers. Advancement in molecular markers integrated with high throughput technology played a vital role in gene mapping, estimation of Genetic Diversity, finding out the evolution and phylogeny of crop, analysis of heterosis, and assessment of diploid/haploid crops and genotyping of cultivars along with Marker Assisted Breeding (MAB)/Marker Assisted Selection (MAS). Now a days, DArT, SSR, SNP, EST-SSR, ISSR, CAPS, SCAR, etc. with high throughput technologies are very exciting markers, which enhances the crop with desired traits and induces tolerance against biotic and abiotic stresses in a short period of time. Though, technologies have gradually enhanced in these newly developed markers, but all these requirements are not yet fulfilled. Hence, appropriate selection of molecular marker are important, which amalgamates some of these desirable characters to achieve the current demand in crop improvement.

REFERENCES

- Akkaya, M.S., Bhagwat, A.A. and Cregan, P.B. (1992). Length polymorphisms of simple sequence repeat DNA in soybean. *Genetics*. 132(4): 1131-1139.
- Badu-Apraku, B., Garcia-Oliveira, A.L., Petrol, C.D., Hearne, S., Adewale, S.A., Gedil, M. (2021). Genetic diversity and population structure of early and extra-early maturing maize germplasm adapted to sub-Saharan Africa. *BMC Plant Biol.* 21, Article96.
- Barcaccia, G. (2010). Molecular Markers for Characterizing and Conserving Crop Plant Germplasm. In: *Molecular Techniques in Crop Improvement*. [Jain, S., Brar, D. (Eds)] Springer, Dordrecht.
- Botstein, D., White, R.L., Skolnick, M. and Davis, R.W. (1980). Construction of a genetic-linkage map in man using restriction fragment length polymorphisms. *American Journal of Human Genetics*. 32: 314-331.
- Chaikam, V., Nair, S.K., Babu, R., Martinez, L., Tejomurtula, J. and Boddupalli, P.M. (2015). Analysis of effectiveness of R1-nj anthocyanin marker for *in vivo* haploid identification in maize and molecular markers for predicting the inhibition of R1-nj expression. *Theoretical and applied genetics*: 128(1): 159-171.
- Chandran, S., Pukalenty, B., Adhimoolam, K., Manickam, D., Sampathrajan, V., Chocklingam, V. and Natesan, S. (2019). Marker-assisted selection to pyramid the opaque-2 ($\alpha 2$) and β -carotene (*crtRB1*) genes in maize. *Frontiers in Genetics*. 10: 859.
- Cheng, C., Motohashi, R., Tsuchimoto, S., Fukuta, Y., Ohtsubo, H. and Ohtsubo, E. (2003). Polyphyletic origin of cultivated rice: based on the interspersed pattern of SINEs. *Molecular Biology and Evolution*. 20(1): 67-75.
- Chu, C.G., Faris, J.D., Friesen, T.L., Xu S.S. (2006). Molecular mapping of hybrid necrosis genes Ne1 and Ne2 in hexaploid wheat using microsatellite markers. *Theor Appl Genet.* 112: 1374-1381.
- Cruz, V.M., Kilian, A. and Dierig, D.A. (2013). Development of DArT marker platforms and genetic diversity assessment of the U.S. collection of the new oilseed crop *lesquerella* and related species. *PLoS one*. 8(5): e64062.
- Das, G., Patra, J. K. and Baek, K.H. (2017). Insight into MAS: A molecular tool for development of stress resistant and quality of rice through gene stacking. *Frontiers in Plant Science*. 8: 985.
- Datta, D., Gupta, Sanjeev, Chaturvedi, S.K. and Nadarajan, N. (2011). *Molecular Markers in Crop Improvement*. Indian Institute of Pulses Research, Kanpur - 208 024
- Daudi, H., Shimelis, H., Mathew, I., Oteng-Frimpong, R., Ojiewo, C., Vršhney, R.K. (2021). Genetic diversity and population structure of groundnut (*Arachis hypogaea* L.) accessions using phenotypic traits and SSR markers: implications for rust resistance breeding. *Genet Resour Crop Evol.* 68: 581-604.
- Diouf, L., Magwanga, R.O., Gong, W., He, S., Pan, Z., *et al.* (2018). QTL mapping of fiber quality and yield-related traits in an intra-specific upland cotton using genotype by sequencing (GBS). *Int. J. Mol. Sci.* 19: 2: 441.
- Dong, W., Liu, J., Yu, J., Wang, L. and Zhou, S. (2012). Highly variable chloroplast markers for evaluating plant phylogeny at low taxonomic levels and for DNA barcoding. *PLoS one*. 7(4): e35071.
- Fazlikhani, L., Keilwagen, J., Kopahnke, D., Deising, H., Ordon, F. and Perovic, D. (2019). High resolution mapping of RphMBR1012 conferring resistance to *Puccinia hordei* in Barley (*Hordeum vulgare* L.). *Frontiers in Plant Science*. 10: 640.
- Gaballah, M.M., Metwally, A.M., Skalicky, M., Hassan, M.M., Brestic, M., El Sabagh, A. and Fayed, A.M. (2021). Genetic diversity of selected rice genotypes under water stress conditions. *Plants*. 10(1): 27.
- Geng, X., Qu, Y., Jia, Y., He, S., Pan, Z., Wang, L. and Du, X. (2021). Assessment of heterosis based on parental genetic distance estimated with SSR and SNP markers in upland cotton (*Gossypium hirsutum* L.). *BMC genomics*. 22: 123.
- Ghobadi, G., Etmian, A., Mehrabi, A.M., Shooshtari, L. (2021). Molecular diversity analysis in hexaploid wheat (*Triticum aestivum* L.) and two *Aegilops* species (*Aegilops crassa* and *Aegilops cylindrica*) using CBDP and SCoT markers. *J. Genet Eng Biotechnol.* 19: 56.
- Gordeeva, E., Shamanin, V., Shoeva, O., Kukoeva, T., Morgounov, A. and Khlestkina, E. (2020). The strategy for marker-assisted breeding of anthocyanin-rich spring bread wheat (*Triticum aestivum* L.) Cultivars in Western Siberia. *Agronomy*. 10(10): 1603.

- Gouda, G., Gupta, M.K., Donde, R., Mohapatra, T., Vadde, R. and Behera, L. (2020). Marker-assisted selection for grain number and yield-related traits of rice (*Oryza sativa* L.). *Physiology and Molecular Biology of Plants*. 26(5): 885-898.
- Griffiths, A., Miller, J., Suzuki, D., Lewontin, R. and Gelbart, W. (2000). Mapping with molecular markers. An Introduction to Genetic Analysis, 7th edition, New York: WH Freeman.
- Grodzicker, T., Williams, J., Sharp, P. and Sambrook, J. (1974, January). Physical mapping of temperature-sensitive mutations of adenoviruses. In Cold Spring Harbor Symposia on Quantitative Biology. 39: 439-446.
- Hinze, L.L., Hulse-Kemp, A.M., Wilson, I.W., Zhu Q.H., Llewellyn, D.J., Taylor, J.M., Stelly, DM. (2017). Diversity analysis of cotton (*Gossypium hirsutum* L.) germplasm using the CottonSNP63K Array. *BMC Plant Biol*. 17: 37.
- Hossain, M.A., Hossen, M.S. and Karim, M.R. (2020). Molecular markers: Indispensable tools for genetic diversity analysis and crop improvement biotechnology. *Int. J. Plant Breed. Crop. Sci*. 7(1): 613-623.
- Huang, X., Kurata, N., Wei, X., Wang, Z.X., Wang, A., Zhao, Q., Zhao, Y., Liu, K., Lu, H., Li, W., Guo, Y., Lu, Y., Zhou, C., Fan, D., Weng, Q., Zhu, C., Huang, T., Zhang, L., Wang, Y., Feng, L., Han, B. (2012). A map of rice genome variation reveals the origin of cultivated rice. *Nature*. 490(7421): 497-501.
- Kasoma, C., Shimelis, H., Laing, M.D, Shayanowako, A.I.T. and Mathew, I. (2020). Revealing the genetic diversity of maize (*Zea mays* L.) populations by phenotypic traits and DArTseq markers for variable resistance to fall armyworm. *Genetic Resources and Crop Evolution*. 68: 243-259.
- Khush G.S., Virmani S.S. (1996). Haploids in plant breeding. In: *In vitro* haploid production in higher plants. Current Plant Science and Biotechnology in Agriculture. [Jain S.M., Sopory S.K., Veilleux R.E. (eds)], 23. Springer, Dordrecht.
- Kilian, A., Wenzl, P., Huttner, E., Carling, J., Xia, L., Blois, H., Caig, V., Heller-Uszynska, K., Jaccoud, D., Hopper, C., Aschenbrenner-Kilian, M., Evers, M., Peng, K., Cayla, C., Hok, P. and Uszynski, G. (2012). Diversity arrays technology: A generic genome profiling technology on open platforms. *Methods in molecular biology* (Clifton, N.J.). 888: 67-89.
- Konieczny, A. and Ausubel, F.M. (1993). A procedure for mapping Arabidopsis mutations using co dominant ecotype specific PCR based markers. *The plant journal*. 4(2): 403-410.
- Kumar, P., Banjarey, P., Malik, R., Tikle, A.N., Verma, R.P.S. (2020). Population structure and diversity assessment of barley (*Hordeum vulgare* L.) Introduction from ICARDA. *Journal of genetics*. 99: 70.
- Lei, M., Ali, M., Jiang, C., Shen, Z., Cai, Y., Yang, P. and Feng, Z. (2020). Marker-assisted selection in a global barley (*Hordeum vulgare* subsp. *vulgare*) collection revealed a unique genetic determinant of the naked barley controlled by the nud locus. *Genetic Resources and Crop Evolution*. 67(2): 273-280.
- Li, C., Dong, Y., Zhao, T., Li, L., Li, C., Yu, E., Mei, L., Daud, M.K., He, Q., Chen, J. and Zhu, S. (2016). Genome-Wide SNP Linkage Mapping and QTL Analysis for Fiber Quality and Yield Traits in the Upland Cotton Recombinant Inbred Lines Population. *Frontiers in Plant Science*. 7: 1356.
- Litt, M. and Luty, J.A. (1989). A hypervariable microsatellite revealed by *in vitro* amplification of a dinucleotide repeat within the cardiac muscle actin gene. *American Journal of Human Genetics*. 44(3): 397-401.
- Melchinger, A.E., Schipprack, W., Wurschum, T., Chen, S. and Technow, F. (2013). Rapid and accurate identification of *in vivo*-induced haploid seeds based on oil content in maize. *Sci Rep*. 3: 2129.
- Mishra, R.K. and Tomar, R.S. (2014). Molecular markers and their application in genetic mapping. *Eur Acad Res*. 2(3): 4012-4040.
- Moreno Gonzalez, J. (1999). Molecular markers and heterosis. *Genetics and Exploitation of Heterosis in Crops*. 257-268.
- Mullis, K. (1990). The unusual origin of the polymerase chain Reaction. *Scientific American*. 262(4): 56-65.
- Nadeem, M.A., Nawaz, M.A., Shahid, M.Q., Doğan, Y., Comertpay, G., et al. (2018). DNA molecular markers in plant breeding: current status and recent advancements in genomic selection and genome editing. *Biotechnology and Biotechnological Equipment*. 32(2): 261-285.
- Naik, N., Rout, P., Umakanta, N., Verma, R.L., Katara, J.L., Sahoo, K.K. and Samantaray, S. (2017). Development of doubled haploids from an elite indica rice hybrid (BS6444G) using anther culture. *Plant Cell, Tissue and Organ Culture (PCTOC)*. 128(3): 679-689.
- Nie, Y., Ji, W. and Ma, S. (2019). Assessment of heterosis based on genetic distance estimated using SNP in common wheat. *Agronomy*. 9(2): 66.
- Noormohammadi, Z., Ibrahim-Khalili, N., Sheidai, M. and Alishah, O. (2018). Genetic fingerprinting of diploid and tetraploid cotton cultivars by retrotransposon-based markers. *The Nucleus*. 61(2): 137-143.
- Oka, H.I. Origin of cultivated rice. (Japan Scientific Societies Press, 1988).
- Olson, M., Hood, L., Cantor, C. and Botstein, D. (1989). A common language for physical mapping of the human genome. *Science* (New York, N.Y.). 245(4925): 1434-1435.
- Pandey, M.K., Gangurde, S.S., Sharma, V., Pattanashetti, S.K., Naidu, G.K., Faye, I., Varshney, R.K. (2021). Improved genetic map identified major QTLs for drought tolerance and Iron deficiency tolerance-related traits in groundnut. *Genes*. 12(1): 37.
- Paran, I. and Michelmore, R.W. (1993). Development of reliable PCR-based markers linked to downy mildew resistance genes in lettuce. *Theoretical and Applied Genetics*. 85(8): 985-993.
- Pradhan, S.K., Pandit, E., Pawar, S., Bharati, B., Chatopadhyay, K., Singh, S., Reddy, J.N. (2019). Association mapping reveals multiple QTLs for grain protein content in rice useful for biofortification. *Mol Genet Genomics*. 294: 963-983.
- Pavani, M., Sundaram, R.M., Ramesha, M.S., Kishor, P.K. and Kemparaju, K.B. (2018). Prediction of heterosis in rice based on divergence of morphological and molecular markers. *Journal of Genetics*. 97(5): 1263-1279.
- Radi, F., Torok, K., Nagymihaly, M., Kereszt, A. and Dudits, D. (2020). Improved reliability in production of maize inbred lines by the combination of the R1-navajo marker with flow cytometry or microsatellite genotyping. *Cereal Research Communications*. 48(4): 423-430.

- Rafalski A. (2002). Applications of single nucleotide polymorphisms in crop genetics. *Current Opinion in Plant Biology*. 5(2): 94-100.
- Rajendran, A., Muthiah, A., Joel, J., Shanmugasundaram, P. and Raju, D. (2014). Heterotic grouping and patterning of quality protein maize inbreds based on genetic and molecular marker studies. *Turkish Journal of Biology*. 38(1): 10-20.
- Ramesh, P., Mallikarjuna, G., Sameena, S., Kumar, A., Gurulakshmi, K., Reddy, B.V., Reddy, P. and Sekhar, A.C. (2020). Advancements in molecular marker technologies and their applications in diversity studies. *Journal of Biosciences*. 45: 123.
- Ramesh, U.M., Methre, R., Kumar, N.V.M., Katageri, I.S., Gowda, S.A., Adiger, S., Lachagari, V.B.R. (2019). Genome mapping and molecular markers identification for yield, yield component and fibre quality traits in tetraploid cotton. *Plant Breeding*. 138(6): 880-896.
- Sansaloni, C., Petroli, C., Jaccoud, D., Carling, J., Detering, F., Grattapaglia, D., Kilian, A. (2011). Diversity Arrays Technology (DArT) and next-generation sequencing combined: genome-wide, high throughput, highly informative genotyping for molecular breeding of Eucalyptus. *BMC Proc*. 5: P54.
- Shang, L., Wang, Y., Wang, X., Liu, F., Abduweli, A., Cai, S., Li, Y., Ma, L., Wang, K. and Hua, J. (2016). Genetic analysis and QTL detection on fiber traits using two recombinant inbred lines and their backcross populations in upland Cotton. *G3 (Bethesda, Md.)*. 6(9): 2717-2724.
- Singh, A.K. (2017). Discovery and Role of Molecular Markers Involved in Gene Mapping, Molecular Breeding, and Genetic Diversity. In: *Plant Bioinformatics*. [Hakeem K., Malik A., Vardar-Sukan F., Ozturk M. (eds)], Springer, Cham.
- Singh B.D., Singh A.K. (2015). Phylogenetic Relationships and Genetic Diversity. In: *Marker-Assisted Plant Breeding: Principles and Practices*. Springer, New Delhi.
- Smith, H.O. and Wilcox, K.W. (1970). A restriction enzyme from *Hemophilus influenzae* L. Purification and general properties. *Journal of Molecular Biology*. 51(2): 379-391.
- Tomkowiak, A., Bocianowski, J., Wolko, Ł., Adamczyk, J., Mikołajczyk, S. and Kowalczewski, P.Ł. (2019). Identification of markers associated with yield traits and morphological features in maize (*Zea mays* L.). *Plants*. 8(9): 330.
- Vos, P., Hogers, R., Bleeker, M., Reijans, M., Lee, T.V.D., Hornes, M. and Zabeau, M. (1995). AFLP: A new technique for DNA fingerprinting. *Nucleic Acids Research*. 23(21): 4407-4414.
- Welsh, J. and McClelland, M. (1990). Fingerprinting genomes using PCR with arbitrary primers. *Nucleic Acids Research*. 18(24): 7213-7218.
- Wenzl, P., Carling, J., Kudrna, D., Jaccoud, D., Huttner, E. and Kleinhofs, A., *et al.* (2004). Diversity Arrays Technology (DArT) for Whole-genome Profiling of Barley. *Proceedings of the National Academy of Sciences of the United States of America*. 101: 9915-9920.
- Williams, J.G., Kubelik, A.R., Livak, K.J., Rafalski, J.A. and Tingey, S.V. (1990). DNA polymorphisms amplified by arbitrary primers are useful as genetic markers. *Nucleic acids research*. 18(22): 6531-6535.
- Zebire, A.D. (2020). Applications of molecular markers in Genetic Diversity Studies of maize. *Nig. J. Biotech*. 37(1): 101-108.
- Zhang, X., Lu, C., Xu, R. and Zhou, M. (2015). Development of molecular markers linked to barley heterosis. *Euphytica*. 203(2): 309-319.
- Zhang, Y., Changhui, F.E.N.G., Shu, B.I.E., Xiaogang, W.A.N.G., ZHANG, J., Songbo, X.I.A. and Hongde, Q.I.N. (2018). Analysis of short fruiting branch gene and Marker-assisted selection with SNP linked to its trait in upland cotton. *Journal of Cotton Research*. 1(1): 1-7.
- Zhang, Y., Fan, C., Chen, Y., Wang, R.R.C., Zhang, X., Han, F. and Hu, Z. (2021). Genome evolution during bread wheat formation unveiled by the distribution dynamics of SSR sequences on chromosomes using FISH. *BMC Genomics*. 22(1): 1-15.