



# Indole Acetic Acid (IAA) Mediated Amelioration of Lead (Pb) Stress- Physiological Indices of Mung Bean [*Vigna radiata* (L.) Wilczek]

Sana Saleem<sup>1</sup>, Ghulam Yasin<sup>1</sup>, Ikram Ul Haq<sup>2</sup>, Adeela Altaf<sup>3</sup>, Khalid Hussain<sup>4</sup>, Khalid Nawaz<sup>4</sup>

10.18805/LR-630

## ABSTRACT

**Background:** Heavy metals have their adverse effects on growth and physiology of plant. Plant growth regulators help in improving the growth and physiological phenomenon in plants. A pot culture experiment was devised to explore the ameliorative potential of Indole Acetic Acid (IAA) for toxicity of rhizospheric lead (Pb) on two varieties of Mung bean [*Vigna radiata* (L.) Wilczek].

**Methods:** Seeds of two varieties i.e., M- 8 and MN-92 were grown in earthen pots filled with sandy loam soil and were arranged under complete randomization. Fifteen days after germination, the lead (Pb) was added @ 10mg/kg and 20mg/kg soil as solution Pb NO<sub>3</sub>. Indole Acetic Acid @100.0mM was foliarly sprayed twice at 15 and 30 days of plants emergence. Physiological parameters i.e., Photosynthetic Rate, Transpiration Rate, Stomatal conductance, Sub Stomatal CO<sub>2</sub> Concentration and biomass production in the form of stem, root and leaf dry weights were determined at the age of physiological maturity for three replicates.

**Result:** By application of IAA, photosynthetic rate reduction was declined from 24.61% to 17.78% under 10mg Pb stress and from 55.54% to 27.35% under 20mg Pb stress. Stomatal conductance reduction was declined from 0.56% to 0.28% under 10mg Pb stress and from 3.37% to 1.68% under 20mg Pb stress. Alleviation of Pb stress by IAA for transpiration rate was non significant. Similarly, the role of IAA for alleviation of Pb stress in term of dry weights of stem, root and leaves were non significant statistically.

**Key words:** Biomass, Indole acetic acid, Lead, Mung, Photosynthetic rate, Stomatal conductance, Transpiration rate.

## INTRODUCTION

Rapid increase in trends of urbanization and industrialization, has lead to more heavy metals in the environment during the past decades (Ashraf *et al.*, 2019). The origins of these heavy metals are natural or anthropogenic activities (Pichtel, 2016). The uses of agricultural fertilizers and pesticides, sewage sludge, mining and smelting of metal ores and fossil burnings are the main sources of heavy metals. Plant growth hormones are the phytochemicals which can compensate the adverse effects of heavy metals on plants. Auxins are the phytohormones which have their effects on different phenomenon of plant life like cell elongation, cell division, cell differentiation and expansion of leaf. Auxins induce antioxidant enzymes activities such as catalase and peroxidase. The major hormone among the auxins which is present in plant is Indole acetic acid (Jutta, 2000). Plant hormones have been reported to increase plant yield (Savita *et al.*, 2011) and stomatal movement (Petri *et al.*, 2010). Plant growth regulators have become commercialized in many countries like EU, USA and Japan due to their positive influence on plant growth and yield (Jahan *et al.*, 2019). Mung bean [*Vigna radiata* (L.) R. Wilczek] has its origin in 1,500 BC in subcontinent and was thereafter introduced to the continents of Asia, Africa, Australia and America. In Pakistan, Mung bean is cultivated on an area of 141,000 hectares with mean annual yield of 93,000 tons (Pakistan Economic Survey, 2012). Keeping in view the importance of mung bean, hazardous effects of heavy metals and role of IAA, the

<sup>1</sup>Department of Botany, Bahauddin Zakariya University, Multan, Pakistan.

<sup>2</sup>Institute of Biotechnology and Genetic Engineering (IBGE) University of Sindh, Jamshoro, Pakistan.

<sup>3</sup>Department of Environmental Sciences, Bahauddin Zakariya University, Multan, Pakistan.

<sup>4</sup>Department of Botany, University of Gujrat, Pakistan.

**Corresponding Author:** Ghulam Yasin, Department of Botany, Bahauddin Zakariya University, Multan, Pakistan.

Email: yasingmn\_bzu@yahoo.com

**How to cite this article:** Saleem, S., Yasin, G., Haq, I.U., Altaf, A., Hussain, K. and Nawaz, K. (2021). Indole Acetic Acid (IAA) Mediated Amelioration of Lead (Pb) Stress- Physiological Indices of Mung Bean [*Vigna radiata* (L.) Wilczek]. Legume Research. 44(10): 1152-1158. DOI: 10.18805/LR-630.

**Submitted:** 13-05-2021 **Accepted:** 26-06-2021 **Online:** 28-06-2021

present experiment was designed with objective to find out the ameliorating potential of IAA for Pb toxicity on mung bean.

## MATERIALS AND METHODS

The experiment was designed to find the ameliorating potential of indole acetic acid (IAA) for stress effects of rhizospheric lead (Pb) toxicity on two mung bean [*Vigna radiata* (L.) Wilczek] varieties. The pot culture experiment was conducted in Bahauddine Zakariya University, Multan Pakistan during 2016-17. Effluents hazards free sandy loam soil was selected for the experiment. Soil analysis revealed

that the Ec of soil was 1.5 mM, pH 6.5 and saturation percentage was 56%. Earthen pots of 30 cm diameter were filled with 6 kg soil after mixing it well. Mung bean seeds of two varieties i-e M-8 and MN-92 varieties were obtained from Ayub Agriculture Research Institute, Faisalabad (Pakistan). Lead nitrate salt and IAA of Sigma Aldrich, Japan were purchased. Pots after filling were irrigated and left till getting of field capacity moisture contents. After germination normal agronomic practices like irrigation and two times pesticide spray of 5% Thiodon were performed. Arrangement of pots was complete randomization by design. To develop the metal stress, amounts of lead nitrate was added after calculation to develop lead @10 and 20 mg/kg soil. Control pots were left without addition of salt. Solutions of IAA (100.0 mM) were sprayed at the age of fifteen and thirty days of age.

Treatment plan consisted of following six treatments.

T<sub>1</sub> = Normal soil (Without addition of metal salts) + Distilled water foliar spray.

T<sub>2</sub> = Normal soil (Without addition of metal salts) + 100.0 mM IAA foliar spray.

T<sub>3</sub> = Lead (10.0 mg/kg soil) + Distilled water foliar spray.

T<sub>4</sub> = Lead (20.0mg/kg soil) + Distilled water foliar spray.

T<sub>5</sub> = Lead (10.0 mg/kg soil) + 100.0 mM IAA foliar spray.

T<sub>6</sub> = Lead (20.0mg/kg soil) + 100.0 mM IAA foliar spray.

There were five pots for each treatment. After thinning, three plants were maintained in each pot. Gas exchange parameters of leaf such as photosynthetic rate (A), internal CO<sub>2</sub> (Ci) concentration, stomatal conductance (gs), water use efficiency (A/E) and transpiration rate (E) were measured from youngest fully expanded leaf specified from the top of each plant which has a mean leaf area of 4.72 cm<sup>2</sup>. For this, an open system LCA-4 ADC portable InfraRed gas analyzer (IRGA) of Analytical Development Company, Hoddesdon, England was used. Timing of measurements were from 11.00 a.m to 1.00 p.m. Followings specifications were adjusted: Molar flow of air was 403.3 mmol m<sup>-2</sup>s<sup>-1</sup> per unit leaf area, 99.9 kPa atmospheric pressure, 6.0 to 8.9 mbar water vapor pressure of chamber, photon flux density at the surface of was 1711 μmol/m<sup>2</sup>/s, leaf temperature range was 28.4-32.4°C and external CO<sub>2</sub> concentration was 370 mmol/ mol.; Biomass production as stem, root and leaf dry weight were

evaluated after 10 days of PGRs spray completion (40 days old plants). Biomass was determined with electrical balance after drying samples for 48 hours at 80°C. The data of experiment were analyzed statistically using software of COSTAT computer package. Keeping significance level of 5%, Duncan's multiple range test was applied comparing mean values (Duncan, 1955). MSTAT-C Computer Statistical Programm was used. Wherever, F values were significant for testing of means using LSD tests.

## RESULTS AND DISCUSSION

The analysis of variance (Table 1) related to photosynthetic rate showed that significant differences were found among different treatments and varieties. Responses of varieties to treatment showed non-significant differences separately. IAA treatment caused an increase in photosynthetic rate (23.18%). Low and high lead doses decreased the photosynthetic rate by a value of 24.61 and 55.54% respectively. IAA ameliorated the lead stress so growth was reduced by a value of 17.78 and 27.35% in 10 and 20 mg lead treated plants (Table 2).

The analysis of variance of transpiration rate (Table 1) showed that treatments differed non significantly and separate response of both variety to each treatment differed non significantly. While significant differences were observed between two varieties. Increase in transpiration rate by the application of IAA was (8.02%). Lead toxicity of low and high levels decreased the transpiration rate 2.87 and 14.12% respectively. IAA ameliorates the lead toxicity and metal effects on transpiration rate were reduced to values of 1.73 and 7.28% respectively (Table 3).

The analysis of variance (Table 1) showed that stomatal conductance significantly affected by different treatments. Results revealed that varieties and treatments were differed highly significantly. Separate responses of varieties to treatments revealed non-significant differences. Stomatal conductance was increased by exogenous IAA spray as 1.96%. Low and high lead treatments decreased the stomatal conductance 0.56 and 3.37% respectively. IAA application decreased the harmful effect of lead therefore effects of metal on stomatal conductance were lowered to values of 0.28 and 1.68% respectively (Table 4).

**Table 1:** Mean sum of squares for photosynthetic rate, transpiration rate, stomatal conductance, sub stomatal CO<sub>2</sub> concentration, Stem dry weight, root dry weight and leaf dry weight of mung bean [*Vigna radiata* (L.) Wickzek].

Source	df	MSS						
		Photosynthetic rate	Transpiration rate	Stomatal conductance	Internal CO <sub>2</sub> concentration	Stem dry weight	Root dry weight	Leaf dry weight
Treatment (T)	5	43.814***	0.234 <sup>ns</sup>	241.577 *	551.977 <sup>ns</sup>	0.924 <sup>ns</sup>	0.254 <sup>ns</sup>	0.204 <sup>ns</sup>
Variety (V)	1	34.251*	3.540*	224.044 ***	0747.111 <sup>ns</sup>	0.511 <sup>ns</sup>	0.138 <sup>ns</sup>	0.012 <sup>ns</sup>
T × V	5	2.387 <sup>ns</sup>	0.004 <sup>ns</sup>	32.177 <sup>ns</sup>	68.311 <sup>ns</sup>	0.007 <sup>ns</sup>	0.001 <sup>ns</sup>	7.427 <sup>ns</sup>
Error	24	4.908	0.720	66.941	260.916	0.405	0.777	0.662
Total	35							

\*\*\*= Highly significant; \*= Significant; ns= Not significant.

**Table 2:** Means for photosynthetic rate [A;  $\mu\text{mol (CO}_2\text{)} \text{ m}^{-2} \text{ sec}^{-1}$ ] of mung bean [*Vigna radiata* (L.) Wilczek] plants grown under lead stresses and exposed to foliar spray of IAA (LSD=3.761).

	No metal +Distilled water spray	No metal +IAA spray	10 mg/kg Pb + Distilled water spray	20 mg/kg Pb + Distilled water spray	10 mg/kg Pb + IAA spray	20 mg/kg Pb + IAA spray	Variety mean (LSD=0.955)
MN-92	9.280±0.860	10.266±2.039	6.833±1.150	4.000±0.953	7.951±3.121	6.500±1.350	7.435±2.339 [b]
% age difference		+10.62	-26.36	-56.89	-14.32	-29.95	
M-8	10.999±4.549	17.700±3.825	8.400±1.084	4.911±1.081	9.061±3.184	8.190±0.397	9.385±3.878 [a]
% age difference		+60.92	-23.63	-55.35	-17.61	-25.53	+26.22
Treatment mean (LSD=1.601)	10.135±3.074 [ab]	12.485±3.663 [a]	7.640±1.334 [b]	4.506±1.015 [c]	8.332±2.343 [b]	7.363±1.272 [b]	9.073±3.170
% age difference		+23.18	-24.61	-55.54	-17.78	-27.35	

**Table 3:** Means for transpiration rate [E;  $\text{mmol (H}_2\text{O)} \text{ m}^{-2} \text{ sec}^{-1}$ ] of mung bean [*Vigna radiata* (L.) Wilczek] plants grown under lead stresses and exposed to foliar spray of IAA (LSD=3.761).

	No metal +Distilled water spray	No metal +IAA spray	10 mg/kg Pb + Distilled water spray	20 mg/kg Pb + Distilled water spray	10 mg/kg Pb + IAA spray	20 mg/kg Pb + IAA spray	Variety mean (LSD=0.955)
MN-92	1.680±0.336	1.910±0.667	1.600±0.419	1.400±0.450	1.664±0.127	1.540±0.670	1.644±0.436 [b]
% age difference		+13.69	-4.76	-16.66	-1.19	-8.33	
M-8	2.350±1.095	2.446±1.385	2.250±0.802	2.000±1.101	2.300±1.374	2.210±1.014	2.271±0.930 [a]
%age difference		+4.08	-4.25	-14.89	-2.12	-5.95	+38.13
Treatment mean (LSD=1.601)	2.018±0.628	2.180±1.014	1.960±0.663	1.733±0.836	1.983±0.939	1.871±0.849	1.985±0.784
% age difference		+8.02	-2.87	-14.12	-1.73	-7.28	

The analysis of variance of sub stomatal CO<sub>2</sub> concentration (Table 1) showed that varieties, treatment response of each variety to treatments differed non significantly. Low and high lead treatments decreased the sub stomatal CO<sub>2</sub> concentration 1.70 and 5.98% respectively. IAA treatment increased the sub stomatal CO<sub>2</sub> concentration (1.70%). IAA alleviated the toxic effect of lead and CO<sub>2</sub> concentration reductions were 0.56 and 3.41% respectively under low and high levels of stresses (Table 5).

Reduction in photosynthetic rate, CO<sub>2</sub> assimilation and transpiration rate might be due to decline in water potential (Atteya, 2002). Water contents in metal treated plants are decreased (Poschenrieder *et al.* 1989) due to increasing resistance in water flow (Barcelo *et al.*, 1988) or alteration of cell wall properties by metal (Poschenrieder *et al.*, 1989).

Conduction and transport of water is influenced in root by toxic metals (Barcelo and Poschenrieder, 1990). Photosynthetic reduction might be due to chlorophyll decrease by metal. Hampp *et al.* (1974) have shown that enzymes of chlorophyll biosynthesis like 6-amino laevulinic acid dehydratase and porphobilinogenase are affected in by metal treatment. Lead is also found to depress the rate of photosynthesis (Carlson *et al.*, 1975). Decrease in activities of many enzymes of CO<sub>2</sub> fixation (Barcelo *et al.*, 1988); changes in the thylakoid structure (Fodor *et al.*, 1996) may contribute to reduction in photosynthetic activity and growth. Reduction in plant growth perhaps is conducive to Reactive oxygen species (ROS) production as heavy metals toxicity as an efficient generator of toxic ROS inhibits photosynthetic ETC (Kappus, 1985).

**Table 4:** Means for stomatal conductance [gs; mmol (CO<sub>2</sub>) m<sup>-2</sup> sec<sup>-1</sup>] of mung bean [*Vigna radiata* (L.) Wickzek] plants grown under lead stresses and exposed to foliar spray of IAA (LSD = 15.57).

	No metal +Distilled water spray	No metal +IAA spray	10 mg/kg Pb + Distilled water spray	20 mg/kg Pb + Distilled water spray	10 mg/kg Pb + IAA spray	20 mg/kg Pb + IAA spray	Variety mean (LSD=0.989)
MN-92	349±6.082	360±8.736	345±8.717	337±3.124	374±3.786	340±24.006	346±7.437 [b]
% age difference		+0.27	-1.14	-3.43	-0.57	-2.57	
M-8	365±1.527	366±1.154	363±1.527	351±1.527	364±1.527	361±1.527	361±10.940 [a]
% age difference		+0.27	-0.54	-3.83	-0.27	-1.09	+4.33
Treatment mean (LSD=1.713)	356±9.786 [ab]	363±6.870 [a]	354±11.178 [abc]	344±8.342 [c]	355±10.014 [ab]	350±19.072 [bc]	354±12.207
% age difference		+1.96	-0.56	-3.37	-0.28	-1.68	

**Table 5:** Means for sub stomatal CO<sub>2</sub> concentration (Ci; μmol. mol<sup>-1</sup>) of mung bean [*Vigna radiata* (L.) Wickzek] plants grown under lead stresses and exposed to foliar spray of IAA (LSD = 27.220).

	No metal +Distilled water spray	No metal +IAA spray	10 mg/kg Pb + Distilled water spray	20 mg/kg Pb + Distilled water spray	10 mg/kg Pb + IAA spray	20 mg/kg Pb + IAA spray	Variety mean (LSD=11.112)
MN-92	349±5.567[ab]	355±10.440	343±16.502	319±22.810	346±1.732	333±20.223	341±17.399
% age difference		+1.71	-1.71	-8.59	-0.85	-4.58	
M-8	355±24.111	360±20.000	348±18.193	340±9.073	352±9.814	346±17.156	350±15.793
% age difference		+1.40	-1.97	-4.22	-0.84	-2.53	+2.63
Treatment mean (LSD=1.713)	351±15.811	357±14.529	345±15.715	330±19.322	349±17.280	339±18.293	345±16.996
% age difference		+1.70	-1.70	-5.98	-0.56	-3.41	

**Table 6:** Means for stem dry weight (g) of mung bean [*Vigna radiata* (L.) Wickzek] plants grown under lead stresses and exposed to foliar spray of IAA (LSD=1.072).

	No metal +Distilled water spray	No metal +IAA spray	10 mg/kg Pb + Distilled water spray	20 mg/kg + Distilled water spray	Pb10 mg/kg Pb + IAA spray	20 mg/kg Pb + IAA spray	Variety mMean (LSD=0.437)
MN-92	2.033±0.970	2.21±0.785	1.473±0.596	1.206±0.663	1.760±0.620	1.513±0.685	1.70±0.701
% age difference		+8.70	-27.54	-40.67	-13.42	-25.57	
M-8	2.313±0.455	2.516±0.484	1.700±0.300	1.366±0.550	2.02±0.670	1.75±0.607	1.946±0.564
% age difference		+8.77	-62.03	-40.94	-12.66	-24.34	+13.93
Treatment mean (LSD=0.758)	2.173±0.695	2.363±0.607	1.586±0.439	1.313±0.548	1.893±0.595	1.635±0.594	1.82±0.632
% age difference		+8.74	-27.01	-39.57	-12.88	-24.75	

**Table 7:** Means for root dry weight (g) of mung bean [*Vigna radiata* (L.) Wilczek] plants grown under lead stresses and exposed to foliar spray of IAA (LSD = 1.810).

	No metal + Distilled water spray	No metal + IAA spray	10 mg/kg Pb + Distilled water spray	20 mg/kg Pb + Distilled water spray	10 mg/kg Pb + IAA spray	20 mg/kg Pb + IAA spray	Variety mean (LSD=0.989)
MN-92	1.822±0.871	1.930±1.000	1.544±0.691	1.392±0.911	1.676±0.794	1.555±0.794	1.644±0.732
% age difference		+5.92	-15.25	-23.60	-8.01	-14.65	
M-8	1.892±1.033	2.111±0.92	1.674±0.911	1.500±0.791	1.811±0.999	1.677±0.792	1.771±0.792
% age difference		+11.57	-11.52	-20.71	-4.28	-11.36	+7.72
Treatment mean (LSD=1.713)	1.862±0.851	2.021±0.862	1.600±0.722	1.441±0.764	1.741±0.800	1.612±0.711	1.711±0.750
% age difference		+8.53	-14.07	-22.61	-6.49	-13.42	

**Table 8:** Means for leaf dry weight (g) of mung bean [*Vigna radiata* (L.) Wilczek] plants grown under lead stresses and exposed to foliar spray of IAA (LSD=1.371).

	No metal + Distilled water spray	No metal + IAA spray	10 mg/kg Pb + Distilled water spray	20 mg/kg Pb + Distilled water spray	10 mg/kg Pb + IAA spray	20 mg/kg Pb + IAA spray	Variety mean (LSD=0.955)
MN-92	1.796±0.800	1.900±0.900	1.546±0.690	1.396±0.850	1.676±0.820	1.546±0.691	1.605±0.720
% age difference		+5.79	-13.91	-22.27	-6.68	-13.91	
M-8	1.766±0.850	1.836±0.901	1.513±0.785	1.330±0.923	1.663±0.765	1.520±0.751	1.642±0.690
% age difference		+3.96	-14.32	-24.68	-5.83	-13.93	+2.30
Treatment mean (LSD=1.601)	1.781±0.738	1.868±0.805	1.530±0.661	1.365±0.794	1.666±0.709	1.533±0.646	1.624±0.695
% age difference		+4.88	-14.09	-23.35	-6.45	-15.92	



The data related to analysis of variance of stem dry weight (Table 1) showed that varieties, treatments and response of individual variety to every treatment differed non significantly. Foliar application of IAA non significantly increased the stem dry weight in two varieties (8.74%). Low and high lead treatments decreased the stem dry weight by values of 27.01 and 39.57% respectively. The lead toxicity was decreased due to foliar application of IAA and dry weight reductions were by values of 12.88 and 24.75% respectively (Table 6).

The analysis of variance of root dry weight (Table 1) showed that treatments, varieties and separate response of varieties to treatment differed non significantly. Application of IAA increased the root dry weight (8.53%). Low and high lead doses decreased the root dry weight 14.07 and 22.61% respectively (Table 7). The analysis of variance of leaf dry weight (Table 1) revealed that varieties, treatments and individual response of all varieties to treatment were different to non significantly degree. Indole Acetic Acid application increased the leaf dry weight upto 4.88% (Table 8). Low and high lead treatments decreased the leaf dry weight 14.09 and 23.35% respectively. Application of IAA alleviated the lead stress. Therefore, leaf dry weight reductions were up to 6.45 and 15.92% respectively (Table 8). Biomass of plant organs decreased due to metal stress (Table 6-8). Such type of findings are reported also by Ouariti *et al.* (1997) and Fengxiang *et al.* (2003). Plant growth and plant water contents have relations with plant weights and biomass. Decline in biomass might be due to Inhibition of both cell elongation and division by heavy metals (Arduini *et al.*, 1994) or due to reduction in nitrogen contents (Andrews *et al.*, 1999). Decrease in dry mass production may also be due to decreased cytokinin as a result of limited nutrients supply (Van der Werf and Nagel, 1996). Reduced amount of cytokinin decreases leaf expansion in plants treated with metal (Gadallah and El-Enany, 1999). Decreased cytokinin contents also affect cell division and cell expansion (Downes and Crowell, 1998). IAA when applied on plants increases net photosynthesis rate and synthesizes more C: N ratio which results in growth enhancement (Sudadi, 2012).

## CONCLUSION

From the results of experiment it can be inferred that foliar application of 100 mM indole acetic acid (IAA) affected the photosynthetic rate and stomatal conductance to statistically significant level. The ameliorative effects of IAA on transpiration rate and internal CO<sub>2</sub> concentration were statistically non significant. Similarly, IAA effects for mitigation of Pb stress in term of dry biomasses were to a non significant level.

## REFERENCES

- Andrews, M., Sprent, J.I., Raven, J.A. and Eady, P.E. (1999). Relationship between shoot to root ratio, growth and leaf soluble protein concentration of *Pisum sativum*, *Phaseolus vulgaris* and *Triticum aestivum* under different nutrient deficiencies. *Plant Cell and Environment*. 22: 949-958.
- Arduini, I., Godbold, D. and Onnis, A. (1994). Cadmium and copper change root growth and morphology of *Pinus pinea* and *Pinus piaster* seedlings. *Physiologia Plantarum*. 92: 675-680.
- Ashraf, S., Ali, Q., Zahir, Z.A., Ashraf, S. and Asghar, H.N. (2019). Phytoremediation: Environmentally sustainable way for reclamation of heavy metal polluted soils. *Ecotoxicology and Environmental Safety*. 174: 714-727.
- Atteya, A.M. (2002). Characterization of growth and water relations in barley during water stress and after rewatering. *Al-Azhar Journal of Pharmaceutical Sciences*. 29: 285-296.
- Barcelo, J. and Poschenrieder, C.H. (1990). Plant water relations as affected by heavy metal stress. *Journal of Plant Nutrition*. 13: 1-37.
- Barcelo, J., Vazquez, M.D. and Poschenrieder, C.H. (1988). Cadmium induced structural and ultrastructural changes in the vascular system of bush bean stems. *Acta Botanica*. 10: 254-261.
- Carlson, R.W., Bazzaz, F.A. and Rolfe, G.L. (1975). The effect of heavy metals on plants. Part 11. Net photosynthesis and transpiration of whole corn and sunflower plants treated with Ph, Cd, Ni and Ti. *Environmental Research*. 10: 113-120.
- Downes, B. and Crowell, D. (1998). Cytokinin regulates the expression of a soybean b-expansin gene by a post-transcriptional mechanism. *Plant Molecular Biology*. 37: 437-444.
- Duncan, D.B. (1955). Multiple range and multiple F-test. *Biometrics*. 11: 1-42.
- Fengxiang, X.B., Han, B., Sridhar, M., David, L. and Monts, S.Y. (2003). Phytoavailability and toxicity of trivalent and hexavalent chromium to *Brassica juncea*. *New Phytologist*. 4: 489-499.
- Fodor, F., Sarvari, E., Lang, F. Szigeti, Z. and Cseh, E. (1996). Effects of Pb and Cd on cucumber depending on the Fe-complex in the culture solution. *Journal of Plant Physiology*. 148: 434-439.
- Gadallah, M.A.A. and El-Enany, A.E. (1999). Role of kinetin in alleviation of copper and zinc toxicity in *Lupinus termis* plants. *Plant Growth Regulator*. 29: 151-160.
- Hamp, R., Kriebitzsch, C. and Ziegler, H. (1974). Effects of lead on enzymes of porphyrin biosynthesis in chloroplasts and erythrocytes. *Naturwissenschaften*. 11: 504.
- Jahan, M.A.H.S., Hossain, A., Teixeira da Silva, J.A., Sabagh, A.E.L., Rashid, M.H. and Barutçular, C. (2019). Effect of Naphthaleneacetic acid on root and plant growth and yield of ten irrigated wheat genotypes. *Pakistan Journal of Botany*. 51: 451-459.
- Jutta, L.M. (2000). Indole-3-butyric acid in plant growth and development. *Plant Growth Regulators*. 32: 219-230.
- Kappus, H. (1985). Lipid Peroxidation: Mechanisms, Analysis, Enzymology and Biological Relevance. In: *Oxidative Stress*. [Sies H, (ed.)], London: Academic Press. Pp, 273-310.
- Ouariti, H. and Ghorbal, M.H. (1997). Responses of bean and tomato plants to cadmium: Growth, mineral nutrition and nitrate reduction. *Plant Physiology and Biochemistry*. 35: 347-354.
- Pakistan Economic Survey. (2012). Agriculture in Survey Report 2012. Islamabad: Ministry of Finance. pp 17-32.

- Petri, J.L., Leite, B.G. and Couto, M. (2010). Effect of growth regulators on 'Gala' Apple fructification. UFPEL - Universidade Federal de Pelotas, C.P. 354, 96010-900.
- Pichtel, J. (2016). Oil and gas production wastewater: soil contamination and pollution prevention. *Applied and Environmental Soil Science*. 2: 79-89.
- Poschenrieder, C., Gunse, B. and Barcelo, J. (1989). Influence of cadmium on water relations, stomatal resistance and abscisic acid content in expanding bean leaves. *Plant Physiology*. 90: 1365-71.
- Savita, G., Singh, P.V. and Maurya, N.J. (2011). Responses of *Pisum sativum* L. to exogenous indole acetic acid application under manganese toxicity. *Bulletin of Environmental Contamination and Toxicology*. 86: 605-609.
- Sudadi, S. (2012). Exogenous application of tryptophan and indole acetic acid (IAA) to induce root nodule formation and increase yield of soybean. *Agricultural Science Research Journal*. 2: 134-139.
- Van der Werf, A., Nagel, O.W. (1996). Carbon allocation to shoots and roots in relation to nitrogen supply is mediated by cytokinins and sucrose: opinion. *Plant Soil*. 185: 21-32.